



The Vertical Migratory Rhythm of Intertidal Microphytobenthos in Sediment Depends on the Light Photoperiod, Intensity, and Spectrum: Evidence for a Positive Effect of Blue Wavelengths

Alexandre Barnett^{1,2*}, Vona Méléder^{1,2}, Christine Dupuy¹ and Johann Lavaud^{1,3}

¹ UMR7266 LIENSs 'Littoral, Environnement et Sociétés', Institut du Littoral et de l'Environnement, CNRS/Université de La Rochelle, La Rochelle, France, ² UPRES EA 2160 MMS 'Mer, Molécules, Santé', Faculté des Sciences et Techniques, Université de Nantes, Nantes, France, ³ UMI3376 Takuvik, CNRS/ULaval, Département de Biologie, Université Laval, Quebec City, QC, Canada

OPEN ACCESS

Edited by:

Dongyan Liu,
East China Normal University, China

Reviewed by:

Guang Gao,
Xiamen University, China
Deyong Sun,
Nanjing University of Information
Science and Technology, China

*Correspondence:

Alexandre Barnett
alexandre.barnett@univ-nantes.fr

Specialty section:

This article was submitted to
Marine Ecosystem Ecology,
a section of the journal
Frontiers in Marine Science

Received: 25 January 2020

Accepted: 18 March 2020

Published: 21 April 2020

Citation:

Barnett A, Méléder V, Dupuy C
and Lavaud J (2020) The Vertical
Migratory Rhythm of Intertidal
Microphytobenthos in Sediment
Depends on the Light Photoperiod,
Intensity, and Spectrum: Evidence
for a Positive Effect of Blue
Wavelengths. *Front. Mar. Sci.* 7:212.
doi: 10.3389/fmars.2020.00212

Estuarine intertidal flats strong biological productivity is mainly based on the activity of benthic microalgae communities or microphytobenthos (MPB), mostly dominated by diatoms. Epipelon is a major MPB growth form comprising motile species, which perform repeated "vertical migration" patterns in the upper sediment layers according to tidal and diurnal cycles with upward migration at the beginning of the daylight emersion and downward migration before immersion starts. Although this fascinating behavior has been extensively studied for more than a century, many of its features remain uncharacterized. Epipelon migratory rhythms are believed to be driven by an endogenous internal clock of unknown nature in combination with diverse environmental stimuli. Among the environmental stimuli impacting on MPB vertical migration, light is probably the most important. Rhythmic changes in surface abundance of natural MPB assemblages were therefore continuously assessed at high frequency by Imaging-PAM fluorimetry in fresh sediment sampled at different seasons, comprising 85 migration profiles from 40 sediment samplings over 2 years, and exposed to different light conditions without any other environmental stimuli (i.e., no tidal-like water flow and stable optimal temperature). In particular, we manipulated (i) the 24-h natural photoperiod MPB that was acclimated to in order to disentangle the tight link between the diurnal and tidal rhythmicity of epipelon migration, and (ii) the light spectrum in order to potentially influence MPB accumulation at the surface of sediment. We found that the migration rhythmicity mapped onto the tidal cycle but that it was modulated, and even overridden, by the diurnal cycle and by the irradiance level during daytime periods with a positive phototactic upward migration up to a certain threshold (in our conditions, 120 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ of white light). Also, we found blue wavelengths (465 nm) triggered

MPB surface accumulation, as compared to other wavelengths (white, green, and red) in patterns that were intensity-dependent and species-dependent. In particular, we found two species, *Navicula spartinetensis* and *Gyrosigma fasciola*, which strongly migrate up under blue light and could potentially be used as model species for further studying the light-responses of intertidal MPB.

Keywords: blue light, diatoms, intertidal flats, pigments, migration, photoperiod

INTRODUCTION

Estuarine intertidal flats are among the most productive ecosystems on Earth (Underwood and Kromkamp, 1999). They play a paramount role in the functioning of coastal food-webs and they support essential local socio-economic activities (Lebreton et al., 2019). Tidal flats high biological productivity is mainly based on the activity of benthic microalgal communities or microphytobenthos (MPB) (Admiraal, 1984; Lebreton et al., 2019). In temperate marine areas, MPB is mostly dominated by diatoms (Ribeiro et al., 2013), which display a high degree of taxonomic and functional diversity (Kooistra et al., 2007; Barnett et al., 2015; Nakov et al., 2018). A major growth form of MPB diatoms is epipelon, which mostly inhabits fine cohesive sediments and comprises raphid diatom species moving freely in between sediment particles. The motility of epipellic diatoms is ensured by a slit (raphe) in their cell wall (frustule), which allows for directional and reversible locomotion (Cohn, 2001; Cohn et al., 2016) driven by highly sophisticated suites of micromovements (Apoya-Horton et al., 2006; Bertrand, 2008). Motility is a key adaptation to the intertidal sedimentary habitat providing epipellic diatoms with many ecological benefits including the colonization of microhabitats with specific light, nutrient, and temperature conditions, coping with environmental extreme variations, optimization of photosynthetic productivity, pheromonal-based sexual attraction, and tidal resuspension avoidance (see Consalvey et al., 2004; Marques Da Silva et al., 2017; Nakov et al., 2018, and references therein). The most known and fascinating display of this motile behavior is the “vertical migration” occurring in the upper sediment layers (Consalvey et al., 2004). Typically, at the beginning of daylight emersion (but not during nighttime), epipellic diatoms move upward and form dense and transient photosynthetic biofilms at the surface of sediment. Then, generally before and/or when immersion and/or night start, they move back downward in the deeper sediment layers. This rhythmic pattern repeats according to tides and photoperiod and is continuously reset to match the daily and fortnightly tide timings and the progressive seasonal changes in day length. Vertical migration was proposed to confer MPB with the ability to optimally exploit the physicochemical gradients of the intertidal sediments for performing photosynthesis and carbon fixation in well-lit upper sediment layers, spatially disconnected from cell division in nutrient-rich, stable, and darkened deeper layers (Saburova and Polikarpov, 2003; Consalvey et al., 2004). In that framework, the acquisition of motility obviously appears as the central motor of pennate diatom adaptation to life,

and dominance, in intertidal sediments (Kooistra et al., 2007; Nakov et al., 2018).

Rhythmic MPB vertical migration has been recognized as a key regulating factor of MPB photosynthetic activity and mudflat primary productivity (Serôdio et al., 2005; Spilmont et al., 2007; Haro et al., 2019). The conceptualization of MPB migration dynamics (Guarini et al., 2008) opened a path to improvements in coupled biological-physical modeling attempts of mudflat primary production (Guarini et al., 2006). In this framework, the quantification of the MPB photosynthetically active biomass (PAB) at the surface of sediment (Herlory et al., 2005) and the precise measurement of its spatial and temporal dynamics (Consalvey et al., 2004), as well as its photosynthetic productivity (Kwon et al., 2012; Cartaxana et al., 2016a; Haro et al., 2019), have received major interest. It is generally accepted that rhythmic MPB migration is first governed by the tidal and diurnal cycles, which, together, control the light availability at the surface of sediment. Therefore, among the numerous environmental stimuli that impact on epipelon motility (i.e., temperature, salinity, nutrient availability, depth of the oxic sediment layer, etc.), light is probably the most important (Consalvey et al., 2004). Intertidal MPB indeed experience an extremely complex and continuously changing light environment (Paterson and Hagerthey, 2001), depending on a multifaceted combination of turbidity of the shallow water column at high tide (HT), and open-field-like light fluctuations at low tide (LT). The sediment-dependent light penetration generates (Kuhl and Jorgensen, 1994; Cartaxana et al., 2016b): (i) a photic zone condensed to the first hundreds of μm , (ii) a deeper light penetration in coarser sediment versus more cohesive fine mud, and (iii) a stronger depth attenuation of the blue band (450–500 nm) of the visible light spectrum and thus a steeper gradient of blue light compared to other spectral bands.

Previous attempts have successfully manipulated the light environment of MPB in controlled laboratory conditions, especially in order to disentangle the impact of the tidal and diurnal cycles on the rhythm of MPB vertical migration (Kwon et al., 2012; Haro et al., 2019). Nevertheless, these studies were performed on permanently submersed intertidal sediment, thereby artificially exacerbating the diurnal versus the tidal response as reported for *in situ* subtidal sediment (Ni Longphuir et al., 2006). Additionally, only one previous work (Wenderoth and Rhiel, 2004) has explored the effect of monochromatic wavelengths on daytime MPB vertical migration. Therefore, the present study is a further complementary step: we used imaging Chl *a* fluorimetry on freshly sampled intertidal sediment to continuously measure at high frequency the changes in surface epipellic PAB without any other abiotic stimuli than light (i.e., in

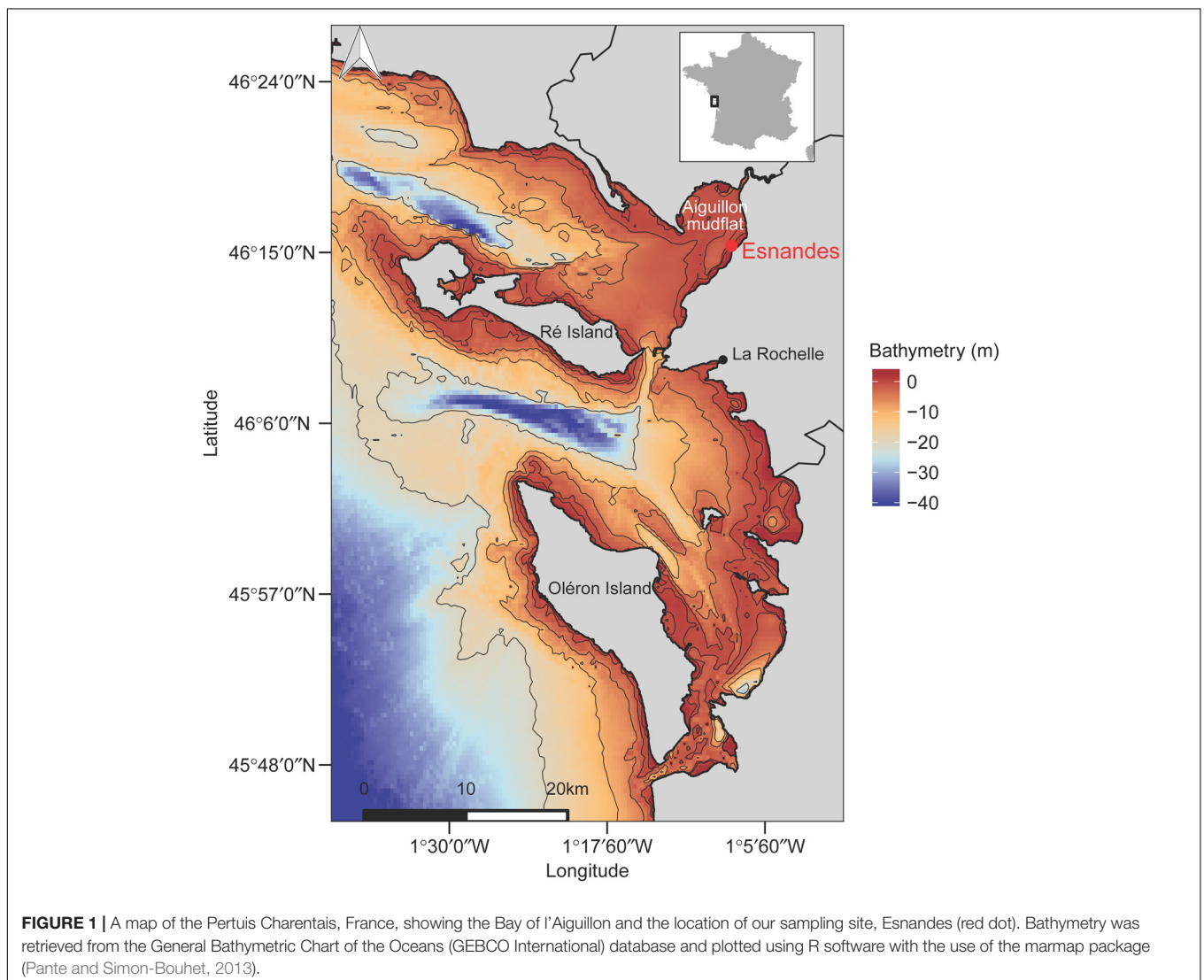
particular, no sediment submersion or tidal-like water flow, and stable optimal temperature). We thereby explored the integrative effects of light photoperiod, intensity, and spectrum at different time scales from the emersion timing to seasonal on the rhythmic changes in MPB abundance at the surface of sediment. We have found that intertidal MPB vertical migration rhythm (i) primarily tracks the tidal and diurnal cycles but to different extents, (ii) is strongly modulated by light intensity and spectrum, (iii) positively influenced by blue wavelengths at the species level by increasing the presence and abundance of cells at the surface of sediment.

MATERIALS AND METHODS

Study Site and Sediment Harvesting

Sediment samples were regularly collected at LT from March 2011 to March 2013 (**Supplementary Table S1**) in the upper zone (500 m from the shore, ca. 4 m above the marine chart

datum) in the bay of l'Aiguillon ("Esnandes" site, $46^{\circ}15.36' N$, $1^{\circ}8.55' W$), an intertidal mudflat site on the Atlantic French coast (**Figure 1**). There, sediment is mostly composed of fine muddy particles (particle size $<63 \mu m$) (Herlory et al., 2004) and is hereafter referred to as "muddy sediment." Occasionally, sediment was also sampled in the bay of Bourgneuf ("Lyarne" site, $47^{\circ}2'35'' N$, $1^{\circ}59'59'' W$) where sediment is composed of muddy sand ($63\text{--}200 \mu m$ particles) (Mélédér et al., 2007) and hereafter referred to as "sandy-muddy sediment." For both sites, the MPB community is largely dominated by diatoms throughout the year (Herlory et al., 2004; Mélédér et al., 2007). The upper layer (approximately the top first centimeter) of sediment was collected at LT and immediately brought back to the laboratory. After sieving through a $500\text{-}\mu m$ mesh to remove macrofauna, especially the mud snail *Peringia* species, the sediment was homogenized by thorough mixing and was spread as a plane layer in 4-cm-depth plastic trays (Serôdio et al., 2012). In order to keep the sediment hydrated throughout the measurements, it was homogenized each 24 h at the time of the *in situ* HT (i.e.,



corresponding to the highest water level at the site of sampling) with filtered (GF/F, 0.7 μm) fresh seawater. In these conditions, MPB conserved its natural migratory rhythm for several days (5 ± 1 days; **Supplementary Figure S1**).

Monitoring of the Motile MPB Abundance Under Different Light Conditions

The “instantaneous” Chl *a* fluorescence F_t was used as a proxy for the MPB biomass present at the surface of sediment (see section “Monitoring of the Changes in the MPB PAB at the Surface of Sediment by Imaging-PAM Fluorimetry”). F_t measurements were converted into Chl *a* concentration per surface of sediment (Chl *a* m^{-2}) as detailed in **Supplementary Figure S2**. F_t was continuously monitored for 24 h on sediment trays placed under an Imaging-PAM fluorometer (Maxi-PAM M-series; Walz, Effeltrich, Germany) and by using its blue (470 nm) detecting beam (settings: intensity 0, frequency 1, measuring frequency 900 s). A map of the irradiance provided by the detecting beam at the surface of sediment tray is shown in **Supplementary Figure S3**. Five “areas of interest” of 6.4 cm^2 , corresponding to 1,500 pixels (Serôdio et al., 2012), and for which the F_t values of all pixels are averaged, were randomly selected. This way, and even if the sediment was well homogenized before each measurement, random “areas of interest” were used as pseudoreplicates for each sediment tray so that each measurement took into account possible remaining heterogeneity of the Chl *a* fluorescence signal across the tray. All experiments were carried out at 20°C, which is optimal for MPB activity throughout the year (Blanchard et al., 1997; Savelli et al., 2018). The reason for maintaining the temperature optimal for all experiments, whatever the season sampling, was to focus only on the effect of light, and therefore to discard effect of temperature on MPB migration that could have potentially confound the output of our experiments.

For each experiment, two phases were recorded: (1) Phase 1: “MPB migration control”, the first 24-h tidal cycle (based on the *in situ* water level at the site of sampling) was recorded under “ambient filtered true light” conditions (i.e., light passing by the window of the laboratory, 24-h natural photoperiod, no direct sunlight) as a control for the changes of the MPB biomass at the surface of the sediment; this option was chosen over an artificial light control in order to take into account the photoperiod and irradiance seasonal changes while keeping the ambient light conditions favorable to MPB migration in the laboratory (i.e., to, for instance, avoid photoinhibitory conditions in summer); (2) Phase 2: after homogenization of the sediment with seawater (see above), the second 24-h tidal cycle was monitored under different conditions of light intensities and spectra using 4 LED panels (FloraLEDs; Plant Climatics, Wertingen, Germany): darkness (Imaging-PAM and sediment tray placed under a black box), “white” light (20, 120, and 350 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$, respectively, the minimum, intermediate, and maximum intensities reached by the mix of different LED sources with 459- and 553-nm maximum wavelengths), blue light (BL; 10, 40, and 120 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$, respectively, the minimum, intermediate, and maximum intensities reached at 465 nm), red

light (180 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$, which was the maximum intensity reached at 677 nm, with and without far red 748 nm), and green light (10 and 40 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$, the minimum and maximum intensities reached by “white” light with a 530-nm filter; 124 Dark Green Lee filter; Lee, Andover, England) (full spectra in **Supplementary Figure S4**). For each light condition, a new sediment tray was used, and a new “MPB migration control” was recorded.

We additionally performed “desynchronization experiments” where the photoperiod was artificially inverted. For doing so, during the “control” Phase 1, the day/night photoperiod was artificially provided following the 24-h natural photoperiod timings by applying a “white light” of 120 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ with the LED panels. During Phase 2, the photoperiod was artificially inverted by applying the same white light during the natural night period and darkness during the natural day period.

The profiles of MPB abundance at the surface of the sediment were analyzed by computing two parameters: (1) the maximum MPB abundance reached during the daytime period of the measurement, (2) the area under the curve that integrates total MPB abundance changes during the 24-h monitoring over the first 24 h of “MPB migration control.” In previous works (Guarini et al., 2006; Coelho et al., 2011), the maximum and integrated MPB abundances were, respectively, used as proxies for the maximum and for the total PAB of MPB present at the surface of the sediment. The graphic analysis of PAB was performed using Excel 2010; total PAB was computed with the Integral (gamma.xla) macro provided by <http://fordom.free.fr>.

For all experiments and under all light conditions described above (**Supplementary Table S2**), we regularly probed the “instantaneous” effective quantum yield of photosystem II (PS II) photochemistry (Φ_{PSIIinst} , see section “Chlorophyll *a* Fluorescence Yield and Light Curves”) under current light conditions and when the daytime maximum MPB abundance was reached:

$$\Phi_{\text{PSIIinst}} = \frac{F'_m - F_t}{F'_m} \quad (1)$$

where F_t is the “instantaneous” Chl *a* fluorescence yield, and F'_m the maximum Chl *a* fluorescence yield measured by firing a saturating pulse (see section “Chlorophyll *a* Fluorescence Yield and Light Curves”).

Sediment Sampling

Sediment cores (diameter 3.2 cm, height 3 cm) were collected in the sediment tray for pigments (see section “Pigments Analysis”) and species diversity (see section “Diatom Species Identification and Diversity Analysis”) analyses. Sampling was performed during the “control” and BL conditions for different seasons: 10 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ BL summer, and 40 and 120 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ BL winter and summer, and at three timings of the corresponding *in situ* tidal cycle: (1) at the timing of the *in situ* LT (i.e., corresponding to the lowest water level at the site of sampling), (2) 3 h before LT (BLT), and (3) 3 h after LT (ALT). The sediment cores were immediately frozen in liquid nitrogen and stored at -80°C .

Pigments Analysis

The pigment content of sediment was determined by reversed phase HPLC (Hitachi High Technologies Co., Tokyo, Japan) calibrated for Chl *a* (expressed in $\mu\text{g Chl } a \text{ cm}^{-2}$; **Supplementary Figure S2**). In order to decipher the vertical pigment content, the surface of three sediment cores (see section “Sediment Sampling”) was cut in three sublayers (0–200, 200–400, and 400–600 μm) with a cryomicrotome (Leica CM1850; Leica Microsystems Nussloch GmbH, Nussloch, Germany). The sediment samples were lyophilized during 24 h, and the pigments were extracted with 0.5 mL of a mixture of 90% methanol/0.2 M ammonium acetate (90/10 vol/vol) and 10% ethyl acetate (Lepetit et al., 2013). Pigment quantification was as described before (Jakob et al., 1999), and Chl *a* content was normalized to the sediment dry weight (g^{-1}). The xanthophyll de-epoxidation state (DES in%) was calculated as:

$$\text{DES} = \frac{\text{DT}}{\text{DD} + \text{DT}} \times 100 \quad (2)$$

where DD is the diadinoxanthin, the epoxidized form; and DT is the diatoxanthin, the de-epoxidized form.

Chlorophyll *a* Fluorescence Yield and Light Curves

In order not to disturb the monitoring of MPB biomass changes with the Imaging-PAM and to discard potential limitations in the measurement of photosynthetic performance by Imaging fluorimetry (Vieira et al., 2013), the Chl *a* fluorescence yield was monitored on the MPB containing sediment surface with a Water-PAM fluorometer (Fiber version; Walz) at the same timings as the sediment sampling, that is, BLT, LT, and ALT (see section “Sediment Sampling”). Chlorophyll *a* fluorescence was excited by a non-actinic modulated 460-nm light, and a saturating pulse of BL ($3,000 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$) was fired in order to, respectively, measure the minimum (F_0) and the maximum (F_m) PS II fluorescence yield on sediment previously dark-adapted for 10 min. The maximum quantum yield of PS II photochemistry was calculated as the ratio F_v/F_m , where

$$F_v = F_m - F_0 \quad (3)$$

Rapid light curves (RLCs) were recorded in triplicates at three different randomly selected locations on the sediment tray. The sediment surface was exposed to eight successive light steps of increasing intensity (E) provided by the LED source of the Water-PAM. The duration of each light step was 30 s as recommended before (Lefebvre et al., 2011). Rapid light curves allowed construction of PSII relative electron transport rate (rETR) versus E curves and non-photochemical quenching (NPQ) versus E curves. Relative electron transport rate was calculated as

$$\text{rETR} = \Phi\text{PS II} \times E \quad (4)$$

Where

$$\Phi\text{PS II} = \frac{F_m' - F'}{F_m'} \quad (5)$$

F' being the fluorescence steady-state yield measured after 30-s illumination at a given E , and F_m' , the maximum fluorescence yield measured at the end of each E by firing a saturating pulse. Non-photochemical quenching was calculated as

$$\text{NPQ} = \frac{F_m - F_m'}{F_m'} \quad (6)$$

NPQ_m was the maximum NPQ induced during the RLC protocol. The rETR- E curves were fitted with the mathematical model of Eilers and Peeters (1988). It allowed extraction of rETR $_m$, the maximum rETR (rETR $_m$, the asymptote of the curve); α , the use light efficiency (the slope at the beginning of the curve); and E_k , the light saturation parameter:

$$E_k = \frac{\text{rETR}_m}{\alpha} \quad (7)$$

Diatom Species Identification and Diversity Analysis

Frozen cores were carefully melted, and “Crème brûlée” samplers (Laviale et al., 2015) were used to collect the first 250 μm of sediment (\emptyset 1.4 cm, 1.5 cm^2). To facilitate cell observation and determination, definitive slides were made after their separation from sediment using Ludox HS-40 colloidal silica (SPCI S.A., St. Denis de la Plaine, France) as described before (Mélédér et al., 2007). A 48-h decantation allowed separation between cells (at the bottom of the tube) and mineral particles (in Ludox). Settled material was rinsed by centrifugation in distilled water (at least five times) and observed with a photonic microscope. Definitive slides were made after cremation (2 h, 450°C) to observe clean diatom cell frustules mounted in a high-resolution diatom mountant (Naphrax; Brunel Microscopes Ltd., Chippenham, Wiltshire, United Kingdom). Species were identified based on morphology (Witkowski, 2000; Ribeiro, 2010). When photonic microscopy was inconclusive, scanning electron microscopy was used. For species composition analysis, a total of ~ 300 diatom frustules were counted to determine specific abundances.

Statistical Analysis

Total and maximum PAB: they were analyzed using one-way analysis of variance (ANOVA) to estimate which PAB proxy was the most impacted by which of the light conditions used. **RLC photosynthetic parameters:** the effects of (i) light conditions (BL 10 and 120 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ and their respective control) and of (ii) the tide sampling timing (BLT, LT, and ALT) were tested using a two-way ANOVA. **Pigment content of sediment:** the effect of the season (Winter and Summer), the light conditions (control winter and summer, and BL 10, 40, and 120 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$), and of the tide sampling timing (BLT, LT, and ALT) were tested using a three-way ANOVA. When significant, ANOVA tests were followed by a *post hoc* Tukey test to highlight differences. **Diatom species abundance:** Hclustering was performed with the Ward.D method in order to group the diatom species and to highlight the groups of interest. All statistical analyses were performed using R software (R Foundation for Statistical Computing, Vienna, Austria).

RESULTS

Vertical Migration Pattern of MPB as a Function of Sediment Type, LT Timing, and Seasons

We monitored MPB abundance at the surface of sediment over 24 h under our light control conditions of 24-h natural photoperiod) at the surface of muddy [dominated by motile epipelon (Seródio et al., 2012) and sandy-muddy sediments (dominated by epipsammon (Méléder et al., 2007)] (Figure 2). For muddy sediment, very low surface MPB abundance occurred at night but rapidly increased with the sun rising, going on through the daytime period until reaching a maximum at approximately solar noon before it decreased (Figure 2A). This bell-like pattern also closely followed the daytime period *in situ* water level at the site of sediment sampling (spring tide), although the sediment was continuously exposed to air in our laboratory conditions. Coincidentally, MPB abundance increased from the day HT to the LT timing, reaching a maximum at a timing close to the one of the lowest *in situ* water level (11:15–12:00 vs. 12:00 h Universal Time), and it then decreased from LT timing toward HT (Figure 2A). These rhythmic changes in MBP abundance were maintained for 5 ± 1 days in our laboratory conditions with, importantly, a sustained daily shift and therefore time correspondence between the timings of LT and of surface MPB maximal abundance (Supplementary Figure S1). Simultaneously, we probed ΦPSII_{inst} , the “instantaneous” effective quantum yield of PS II photochemistry, when the maximum MPB abundance was reached to check if our laboratory conditions negatively influenced photosynthetic potential of MPB and therefore its light response, which was confirmed as not being the case with ΦPSII_{inst} remaining at near optimal levels of 0.689 ± 0.004 . Such verification of ΦPSII_{inst} was subsequently performed for all experiments under all light conditions tested (Supplementary Table S2). For sandy-muddy sediment, we observed a similar (i) coincidence between the maximum MPB surface abundance with solar noon and (ii) inverse relationship between MPB abundance changes and *in situ* water level with a maximum MPB abundance occurring exactly at LT timing (Figure 2B). Nevertheless, major differences appeared compared to muddy sediment: (1) integrated MPB abundance was higher at night, (2) MPB maximum abundance was approximately five times lower during daytime emersion, (3) the speed to reach the MPB maximum abundance was much slower, and (4) after reaching its maximum, MPB abundance remained mostly stable, thus breaking the inverse relationship with the *in situ* water level. ΦPSII_{inst} again remained high (0.666 ± 0.004 , Supplementary Table S2).

Because it provided the strongest and clearest signal, we went on studying migration phenomenon with muddy sediments only. When the *in situ* LT timing did not coincide with solar noon (as in Figure 2A), the pattern of MPB abundance changes was different. Two main patterns complementary to Figure 2A are shown in Supplementary Figure S5 (Supplementary Figure S5A: neap tide, Supplementary Figure S5B: spring tide). When *in situ* LT timing (18:07 h UT) was close to sunset (19:15 h UT), MPB

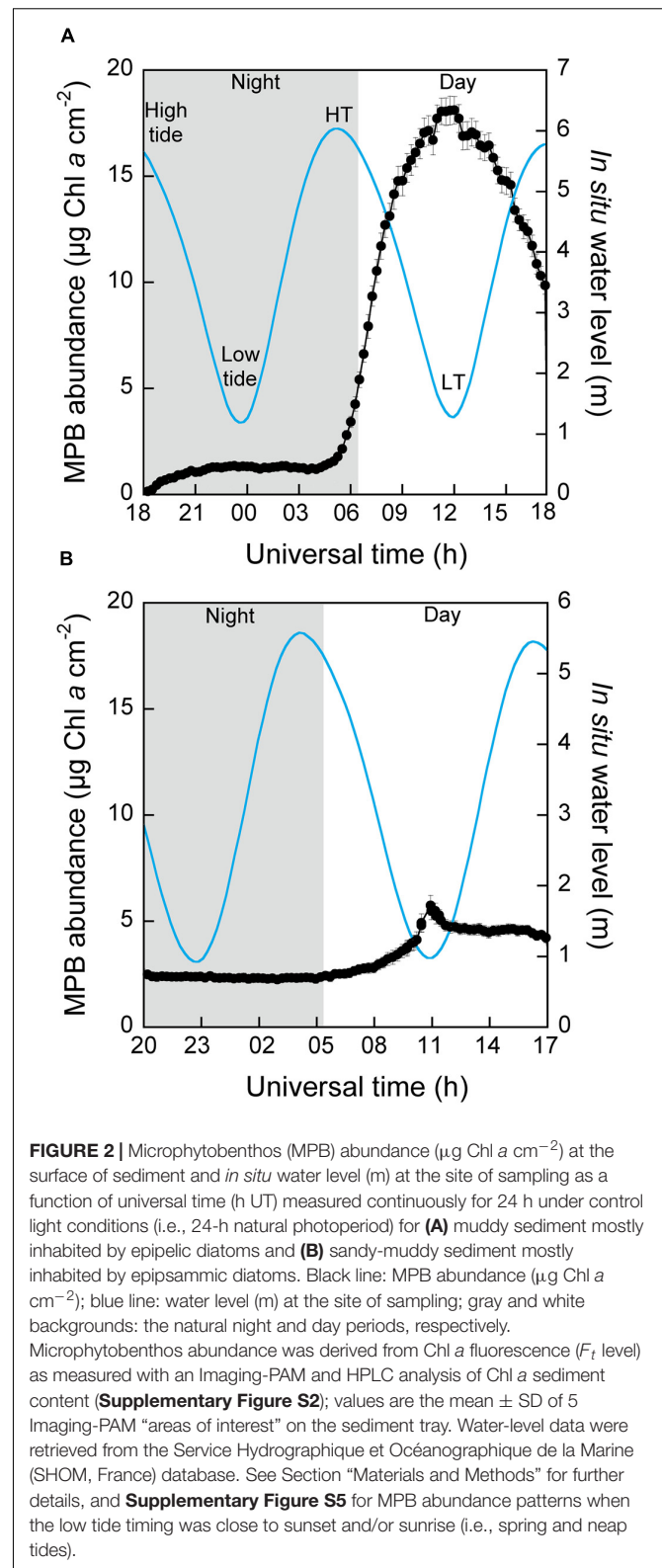


FIGURE 2 | Microphytobenthos (MPB) abundance ($\mu\text{g Chl a cm}^{-2}$) at the surface of sediment and *in situ* water level (m) at the site of sampling as a function of universal time (h UT) measured continuously for 24 h under control light conditions (i.e., 24-h natural photoperiod) for (A) muddy sediment mostly inhabited by epipellic diatoms and (B) sandy-muddy sediment mostly inhabited by epipsammonic diatoms. Black line: MPB abundance ($\mu\text{g Chl a cm}^{-2}$); blue line: water level (m) at the site of sampling; gray and white backgrounds: the natural night and day periods, respectively. Microphytobenthos abundance was derived from Chl a fluorescence (F_t level) as measured with an Imaging-PAM and HPLC analysis of Chl a sediment content (Supplementary Figure S2); values are the mean \pm SD of 5 Imaging-PAM “areas of interest” on the sediment tray. Water-level data were retrieved from the Service Hydrographique et Océanographique de la Marine (SHOM, France) database. See Section “Materials and Methods” for further details, and Supplementary Figure S5 for MPB abundance patterns when the low tide timing was close to sunset and/or sunrise (i.e., spring and neap tides).

abundance started to increase just after the timing of *in situ* HT, again reaching a maximum significantly before (16:55 h UT) LT timing and sunset, so that MPB abundance reached an

approximately 0 value only 3 h after sunset (Supplementary Figure S5A). When *in situ* LT timing was close to sunrise (~ 1 h 15 after sunrise), the maximum MBP abundance followed two patterns as a function of the preceding *in situ* LT timing event (i.e., neap tide or spring tide, respectively): (1) when the later was just before sunset (~ 1 h 15), MPB abundance started to increase the same way as during the preceding daytime period, that is, following the *in situ* water level decrease independent of the day–night alternation (here ~ 2 h before sunrise) and reaching its maximum just after the *in situ* LT timing with no coincidence with solar noon (Supplementary Figure S5A); (2) when the preceding *in situ* LT was at night, MPB abundance started increasing with sunrise, reaching its maximum after the *in situ* LT timing and at a time closer to solar noon (Supplementary Figure S5B).

Taking into account these patterns and their variations, we compiled 85 profiles from 40 sediment samplings over 2 years (Supplementary Table S1) in order to compute (i) the total PAB (Guarini et al., 2006; Coelho et al., 2011) of MPB present at the surface of sediment (the integrated area under the curve), and (ii) the maximum surface PAB reached during daytime LT period. Note that total PAB was computed over 24 h, including nighttime, in order to analyze both MPB abundance changes occurring during daytime (Figure 2A) and closer to sunrise and/or sunset (Supplementary Figure S5). When PAB was plotted versus the *in situ* level of low waters, there was no clear trend (Figure 3A and Supplementary Figure S6A). When plotted versus the *in situ* LT timing, a clearer pattern was observed (Figure 3B and Supplementary Figure S6B): (i) the maximum and total PAB were recorded at daytime LT timings independent of the season; (ii) there was a tendency for the highest values of maximum PAB to be reached when the LT timing coincided with solar noon (ca. $12:00 \pm 2:00$ h UT) as in Figure 2A. When plotted versus sampling date (Figure 3C and Supplementary Figure S6C), the highest PAB values occurred by the end of winter–beginning of spring, they decreased to reach a minimum in summer and autumn and increased again through the next winter. For all experiments, ΦPSII_{inst} was high (0.681 ± 0.026 , Supplementary Table S2).

Vertical Migration Pattern of MPB as a Function of Light Regimes

In a following step, we manipulated the 24-h natural photoperiod to study its impacts on MPB abundance changes at the surface of sediment. First, we artificially replaced the daytime period by night resulting in 24-h continuous darkness and the other way around (Figures 4A,B, respectively). Before such manipulation, MPB migration was recorded for 24 h the same way as described in Figure 2A, reaching a similar maximum PAB: $15.8 \pm 0.7 \mu\text{g Chl } a \text{ cm}^{-2}$ (Figure 4A) and $16.0 \pm 0.7 \mu\text{g Chl } a \text{ cm}^{-2}$ (Figure 4B). When exposed to 24-h darkness, changes in MPB abundance followed a similar pattern as during the preceding 24 h of 24-h natural photoperiod, except the maximum PAB reached during subjective daytime period was lower: $11.3 \pm 0.9 \mu\text{g Chl } a \text{ cm}^{-2}$ (Figure 4A); ΦPSII_{inst} remained high (Supplementary Table S2). When exposed to

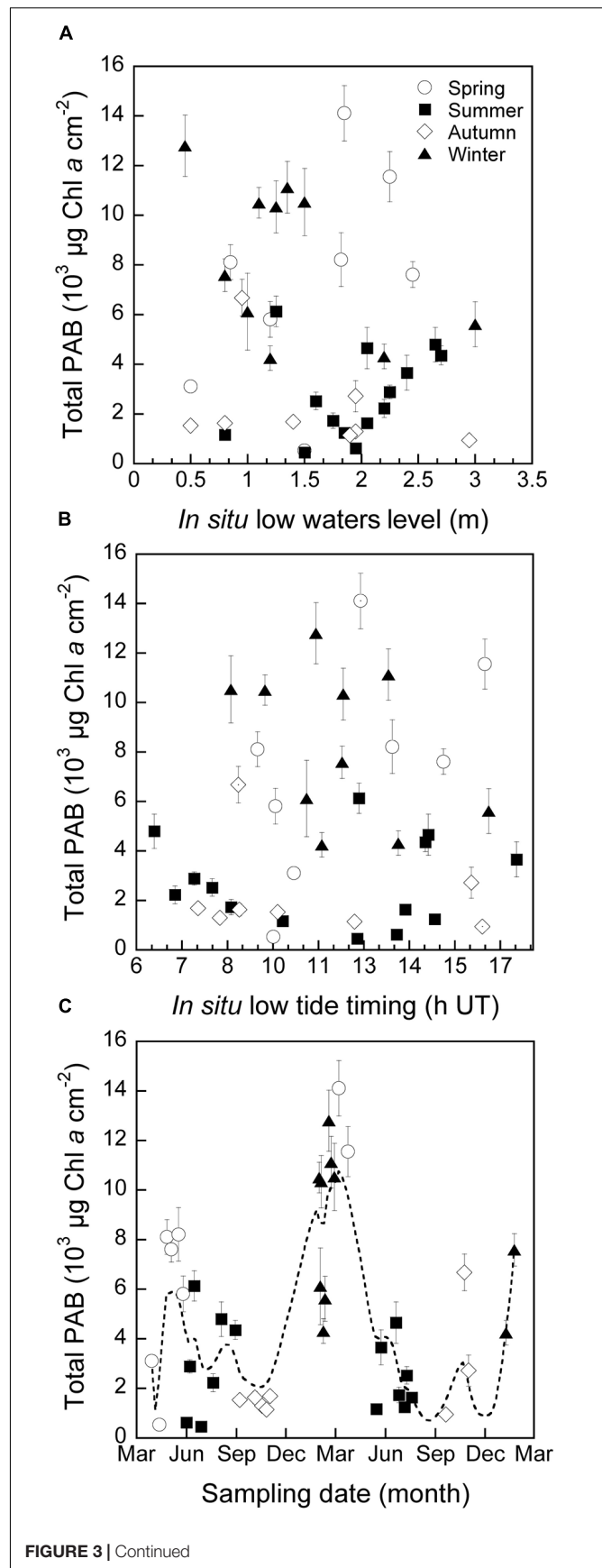


FIGURE 3 | Continued

FIGURE 3 | Microphytobenthos total photosynthetically active biomass (PAB) at the surface of muddy sediment as function of (A) low water level at the site of sampling (i.e., the lowest water level, which depends on the tidal amplitude), (B) the low tide timing (h UT) at the site of sampling (i.e., corresponding to the lowest water level), and (C) the date of sampling (month abbreviations are Mar, March; Jun, June; Sep, September; Dec, December). In order to facilitate visualization, months were pooled under season labels: spring (circles), summer (squares), autumn (diamonds), and winter (triangles). Values are the mean \pm SD of 5 Imaging-PAM “areas of interest” on the sediment tray. Water level and low tide timing data were retrieved from the Service Hydrographique et Océanographique de la Marine (SHOM, France) database.

24-h light (Figure 4B), MPB abundance immediately increased upon onset of the artificial light even if during subjective nighttime period, reaching a maximum ($5.9 \pm 0.6 \mu\text{g Chl } a \text{ cm}^{-2}$) at a time coinciding with the *in situ* LT timing and maintaining at this maximum. During the following daytime period, (i) the increase in MPB abundance was slower than during the first 24-h daytime period so that the occurrence of the maximal PAB was delayed by approximately 2 h (i.e., it occurred 90 min after the *in situ* LT timing instead of 30 min before), and (ii) the value of maximum PAB was much higher: $24.7 \pm 0.6 \mu\text{g Chl } a \text{ cm}^{-2}$. Although ΦPSII_{inst} decreased, it remained relatively high (0.588 ± 0.037 , Supplementary Table S2).

In a second manipulation, we aimed at desynchronizing MPB migration from the 24-h natural photoperiod and eventually from the *in situ* tidal cycle. We continuously monitored MPB migration over several days, and after a first 24-h exposure to an artificial light regime exactly matching the 24-h natural photoperiod, we inverted it by exposing the sediment to darkness during subjective daytime (starting Day 2) and by applying the same artificial light as in Figure 4B during subjective nighttime periods (Figure 4C). After 1 day under the new light regime (Days 2–3 in Figure 4C), MPB abundance changes showed a similar pattern as for the short-time experiments described in Figures 4A,B. This pattern was kept along the 4 following days with a maximum PAB always reached by the end of the light period would it be during artificial daytime (Day 1, similar as Figure 4B, and coinciding with the timing of the *in situ* LT) or subjective nighttime (the following days). Of paramount interest was the fact that in the meantime the timings of the *in situ* LT shifted by 60 ± 15 min per day. It generated a complete loss of synchronization between the timing of maximum PAB with the timing of the *in situ* LT (compare with Supplementary Figure S1), so that by the end of the experiment the maximum PAB was reached at a time nearly coinciding with the timing of the *in situ* HT (03:00 vs. 04:20, Day 6, Figure 4C). ΦPSII_{inst} was 0.648 ± 0.008 for Day 1 and 0.671 ± 0.009 for the following days (Supplementary Table S2).

Vertical Migration Pattern of MPB as a Function of Light Intensity and Light Spectrum

We furthermore tested the effect of light intensities and wavelengths on the PAB at surface of sediment at all seasons,

through experiments with the same light conditions performed at different seasons, thus taking into account seasonal changes in MPB abundance (Figure 3C). Results are compiled in Figure 5, Supplementary Figure S7, and Supplementary Table S3. In order to compare all light conditions, data were normalized on the control light conditions, which were systematically performed before each light treatment (i.e., the first 24-h MPB migration under 24-h natural photoperiod as in Figure 2A). Note that (i) we worked at equivalent PAR (“photosynthetically active radiation”); for technical reasons, we could not work at equivalent PUR (“photosynthetically usable radiation”); (ii) due to technical limitation of our LED panels, we could not use the same PAR for all wavelengths; in particular, monochromatic wavelengths did not reach the highest “white” light PAR ($350 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$); (iii) the statistical analysis was significant only when considering total PAB (one-way ANOVAs, $p \leq 0.05$, Supplementary Table S3) rather than maximum PAB (one-way ANOVAs, $p > 0.05$, Supplementary Table S3). Based on Tukey *post hoc* test, we could draw the following observations. Among all light treatments: (i) a significantly lower total PAB was observed in sustained darkness [$39\% \pm 2\%$ of the control ($p \leq 0.05$) compared to $120 \mu\text{mol photon m}^{-2} \text{ s}^{-1}$ white light and BL and to $40 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$ green light; see Figure 4A and Supplementary Table S3]; (ii) the significantly ($p \leq 0.05$) highest total PAB values were reached for lights of $120 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$ (“white” and blue, $144\% \pm 3\%$ and $176\% \pm 38\%$ of the control, respectively). Yet, lower or higher irradiances generated no significant changes or a significant decrease in total PAB independent of the wavelength (Supplementary Table S3). ΦPSII_{inst} was high under all light conditions: 0.642 ± 0.047 (Supplementary Table S2).

More precisely (Figure 5 and Supplementary Table S3), under “white” light, total PAB significantly increased with irradiance (from 20 to $120 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$) and decreased thereafter (from 120 to $350 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$), so that total PABs under 20 and $350 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$ were not significantly different. Under BL (465 nm), total PAB significantly increased with intensity (from 10 to $120 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$); noteworthy is the effect of an irradiance as low as $10 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$ of BL, which generated a total PAB as high as $75\% \pm 2.77\%$ of the control. Green light (530 nm) did not trigger a response significantly different from the control light conditions ($40 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$) and darkness ($10 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$). Unfortunately, with our setup, we could not test higher green light intensities. The maximum intensity of red light (677 nm, with and without far red 748 nm, $180 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$) did not trigger a total PAB significantly different from the control light.

Effect of BL on the MPB Pigment Content, Photosynthetic Performance, and Diatom Species Composition During Vertical Migration

Because we observed a specific BL positive effect, we analyzed potential changes in (i) the pigment content, (ii) the photosynthetic performance, and (iii) the species composition

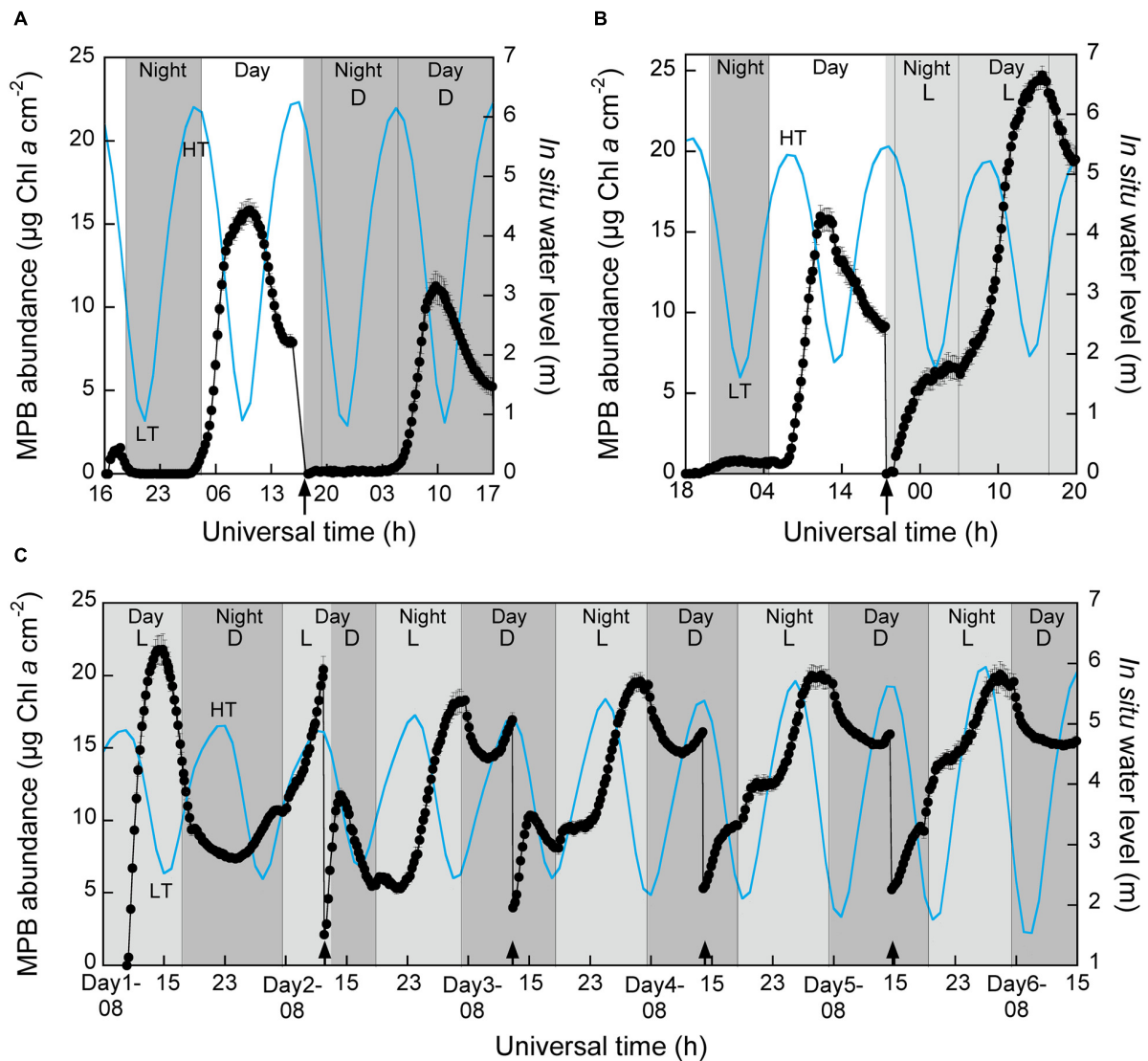


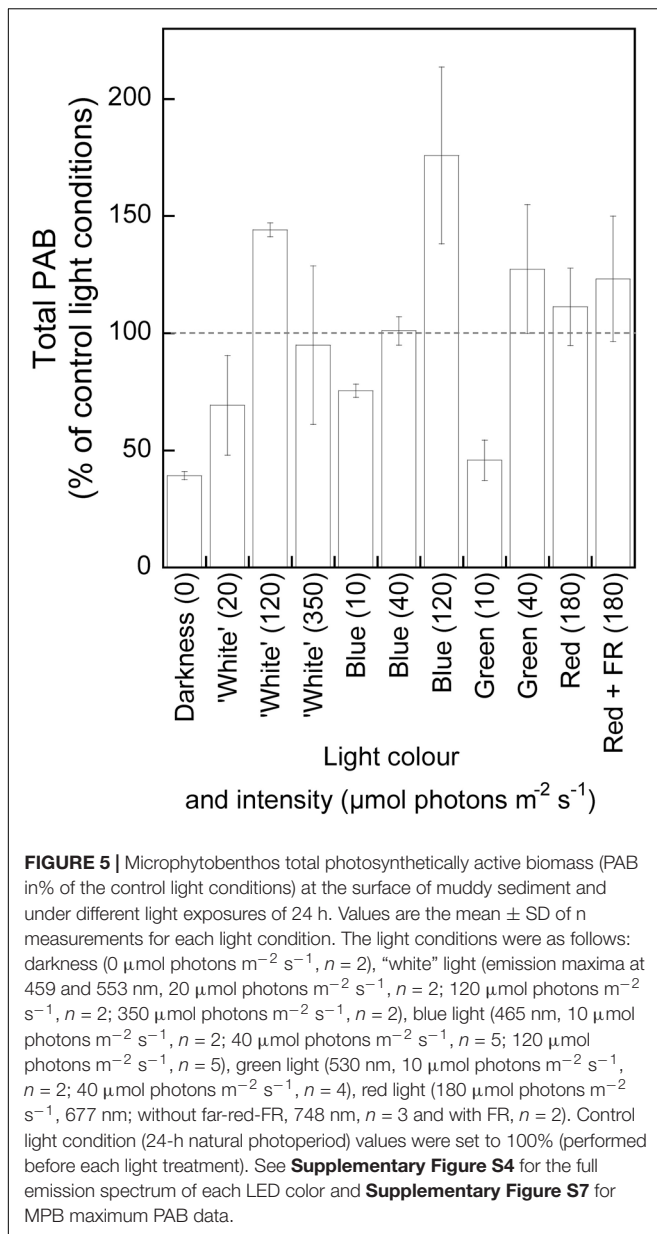
FIGURE 4 | Microphytobenthos abundance ($\mu\text{g Chl } a \text{ cm}^{-2}$) at the surface of muddy sediment, and *in situ* water level (m) at the site of sampling as function of universal time (h UT), and monitored under different light regimes: **(A)** control light conditions for the first natural night/day period ("Night" and "Day" backgrounds) followed by artificial darkness (D) for the second night/day period; **(B)** control light conditions for the first natural night/day period followed by continuous artificial "white" light (L, $120 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$) for the second night/day period, **(C)** artificial "white" light (L) followed by artificial darkness (D) for the first night/day period (Day 1), followed by artificially inverted night/day period (starting Day 2) for the four following days (Days 3–6). Low tide and HT, low tide and HT *in situ* timings, respectively, as in **Figure 2**. Black line: MPB abundance ($\mu\text{g Chl } a \text{ cm}^{-2}$); blue line: water level (m) at the site of sampling; dark gray and light gray backgrounds: the night and day periods, respectively; arrows indicate the timings for which the sediment was stirred with fresh seawater. Microphytobenthos abundance was derived from Chl *a* fluorescence (F_t level) as measured with an Imaging-PAM and HPLC analysis of Chl *a* sediment content (**Supplementary Figure S2**); values are the mean \pm SD of 5 Imaging-PAM "areas of interest" on the sediment tray. Water-level data were retrieved from the Service Hydrographique et Océanographique de la Marine (SHOM, France) database. See the Section "Materials and Methods" for further details.

of MPB during its migration [at the *in situ* LT timing, 3 h before (BLT) and after (ALT)] under BL exposure of different intensities (10 , 40 , and $120 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$) and for winter and summer sediments, when total PAB was the highest and the lowest, respectively (**Figure 3C**).

Pigment Content and Photosynthetic Performance

Pigments were analyzed at three depths (0 – 200 , 200 – 400 , and 400 – $600 \mu\text{m}$; **Supplementary Table S4**) and expressed relative to

Chl *a* in order to normalize for pigment changes due to changes in MPB abundance with seasons and/or tidal cycle (**Figure 3**); $600 \pm 100 \mu\text{m}$ is usually the depth of the photic zone in fine sediments, and 200 – $250 \mu\text{m}$ is where photoprotection is the highest (Cartaxana et al., 2011). We found significant differences in winter versus summer sediments exposed to light control conditions and for most sampling times (LT, BLT, and ALT), as well as some significant differences under BL versus control light conditions (detailed description, **Supplementary Table S4**).



However, we observed no changes in MPB pigment content, and especially in BL absorbing carotenoids, to relate with the BL positive effect on vertical MPB migration described above (see section "Vertical Migration Pattern of MPB as a Function of Light Intensity and Light Spectrum").

Because most of the significant differences in pigment content were observed for sediment sampled in summer (**Supplementary Table S4**), we analyzed MPB photosynthetic performances in summer only and for the BL lowest and highest intensities (10 and 120 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$) (**Table 1**). F_v/F_m (which is equivalent to ΦPSII_{inst} after dark acclimation of MPB) was high and stable for all BL treatments and their respective controls (two-way ANOVA, $p > 0.05$ for all factors, **Table 1**). Because they could not be monitored during a restricted timeframe (i.e.,

two consecutive weeks), the two controls significantly differed in $r\text{ETR}_m$ and E_k , both parameters of the control for BL-10 (sampling end of May) being lower than the control for BL-120 (sampling end of July) (two-way ANOVA, $p \leq 0.001$ for the light effect, different groups with the Tukey test, **Table 1**). While exposure to BL-10 μmol did not yield any significant changes in photosynthetic performance (similar group with Tukey test; **Table 1**), BL-120 generated a significant decrease in $r\text{ETR}_m$ and E_k for all *in situ* tide timings (different group with Tukey test; **Table 1**) but no significant changes in α (two-way ANOVA, $p > 0.05$ for all factors). There was also a tendency for higher NPQ_m values under BL-120 LT versus control conditions (two-way ANOVA, $p \leq 0.05$ for the light effect).

Diatom Species Composition

The MPB species composition was analyzed in the 0- to 250- μm sediment layer. Forty-nine species were identified including a large majority of pennate diatoms and of epipelagic representatives (**Supplementary Table S5**). Because only two species, *Navicula phyllepta* and *Navicula spartinetensis*, were present in all samples, we first proceeded with an Hcluster analysis showing that most of the species (left cluster, **Supplementary Figure S8A**) have a very low abundance. In a second step (**Supplementary Figure S8B**), we proceeded with the 19 species that were present at least once $>5\%$ of the total MPB relative abundance. This way, two species, *N. spartinetensis* and *G. fasciola*, were highlighted. The species relative abundance of all experiments (**Figure 6**) showed that (i) only the exposure to BL-40 generated major changes (**Figures 6B,D**), and (ii) these changes concerned *N. spartinetensis* in summer (**Figure 6B**, pale blue) and *G. fasciola* in winter (**Figure 6D**, yellow). In order to discard a potential effect of changes in the relative abundance of other motile species on *N. spartinetensis* and *G. fasciola* (i.e., for instance their relative abundance would be artificially increased by a decrease in the relative abundance of other motile species), we normalized the relative abundance of the 12 motile species (among the selected 19) encountered in the light conditions of interest (summer and winter, BL-40) to the relative abundance of non-motile species as a common and stable basis: if the abundance of a given species would increase relative to non-motile species, we would assume an upward migration and *vice versa*. This analysis (**Supplementary Figure S9**) confirmed that *N. spartinetensis* and *G. fasciola* were indeed the two species whose abundances increased relative to the pool of non-motile species under BL and that the response of *N. spartinetensis* was particularly strong.

More precisely, for both species, BL induced an increase in abundance at the surface of sediment, all along the LT period (**Supplementary Figure S10**). *Gyrosigma fasciola* was massively present at the surface of sediment (50% of the total abundance of 19 most present species) already by the beginning of LT (BLT) while appearing only by the end of the LT (ALT) under control light conditions (**Supplementary Figure S10B**). Noteworthy were the observations highlighting a respective specific effect of a certain BL dose in the two species: (i) while *N. spartinetensis* abundance continuously increased to reach a maximum of approximately 85% relative abundance at ALT timing, *G. fasciola* abundance decreased by approximately

TABLE 1 | Photosynthetic parameters of microphytobenthos (MPB) exposed to blue (465 nm) light conditions (the blue 10 and 120 $\mu\text{mol photon m}^{-2} \text{s}^{-1}$ intensities between brackets) and their respective control light conditions: because the two blue light exposures could not be performed simultaneously, it was necessary to perform measurements under control light conditions independently for each of them.

		Control	Blue (10)	Control	Blue (120)
BLT	F_v/F_m	0.74 ± 0.02	0.72 ± 0.05	0.74 ± 0.03	0.72 ± 0.03
	α	0.63 ± 0.06	0.63 ± 0.05	0.63 ± 0.05	0.62 ± 0.06
	rETR _m	38.78 ± 6.96 ^{bc}	40.35 ± 7.58 ^{bc}	92.3 ± 19.71 ^a	54.25 ± 11.14 ^b
	E_k	66.58 ± 10.43 ^{bcd}	65.53 ± 16.36 ^{bcd}	146.21 ± 26.08 ^a	88.29 ± 18.45 ^b
	NPQ _m	1.11 ± 0.12 ^{ab}	1.05 ± 0.20 ^{ab}	1.01 ± 0.19 ^{ab}	1.11 ± 0.48 ^{ab}
LT	F_v/F_m	0.75 ± 0.02	0.74 ± 0.02	0.72 ± 0.02	0.74 ± 0.03
	α	0.63 ± 0.05	0.64 ± 0.05	0.63 ± 0.05	0.63 ± 0.07
	rETR _m	36.32 ± 6.48 ^{bc}	36.44 ± 7.93 ^{bc}	91.7 ± 15.18 ^a	45.63 ± 11.78 ^{bc}
	E_k	60.77 ± 9.47 ^{bcd}	58.54 ± 16.9 ^{cd}	147.4 ± 20.05 ^a	67.65 ± 11.47 ^{bcd}
	NPQ _m	1.05 ± 0.18 ^{ab}	1.06 ± 0.26 ^{ab}	1.08 ± 0.26 ^{ab}	1.45 ± 0.46 ^a
ALT	F_v/F_m	0.73 ± 0.03	0.74 ± 0.02	0.71 ± 0.02	0.72 ± 0.07
	α	0.61 ± 0.06	0.64 ± 0.06	0.68 ± 0.07	0.58 ± 0.07
	rETR _m	29.93 ± 8.60 ^c	35.72 ± 8.58 ^{bc}	81.82 ± 20.56 ^a	47.53 ± 3.41 ^{bc}
	E_k	49.55 ± 10.11 ^d	58.23 ± 15.19 ^{cd}	129.02 ± 25.22 ^a	87.35 ± 18.53 ^{bc}
	NPQ _m	0.93 ± 0.17 ^b	0.99 ± 0.24 ^b	1.10 ± 0.22 ^{ab}	1.29 ± 0.35 ^{ab}

Two-way analysis of variance were performed to test the effect of light conditions and of the tide sampling timing. Post hoc Tukey test was further performed to group values. For each photosynthetic parameter, values with the same letter are not significantly different. F_v/F_m , maximum quantum yield of PSII photochemistry (no units); α , maximum light efficiency use (relative units); rETR_m, maximum relative photosynthetic electron transport rate ($\mu\text{mol electrons m}^{-2} \text{s}^{-1}$); E_k , light saturation coefficient ($\mu\text{mol photons m}^{-2} \text{s}^{-1}$); NPQ_m, maximum NPQ (no units); BLT, 3 h before low tide; LT, low tide timing at the site of sampling (i.e., corresponding to the lowest water level); ALT, 3 h after low tide. Values are the mean ± SD of 3 measurements.

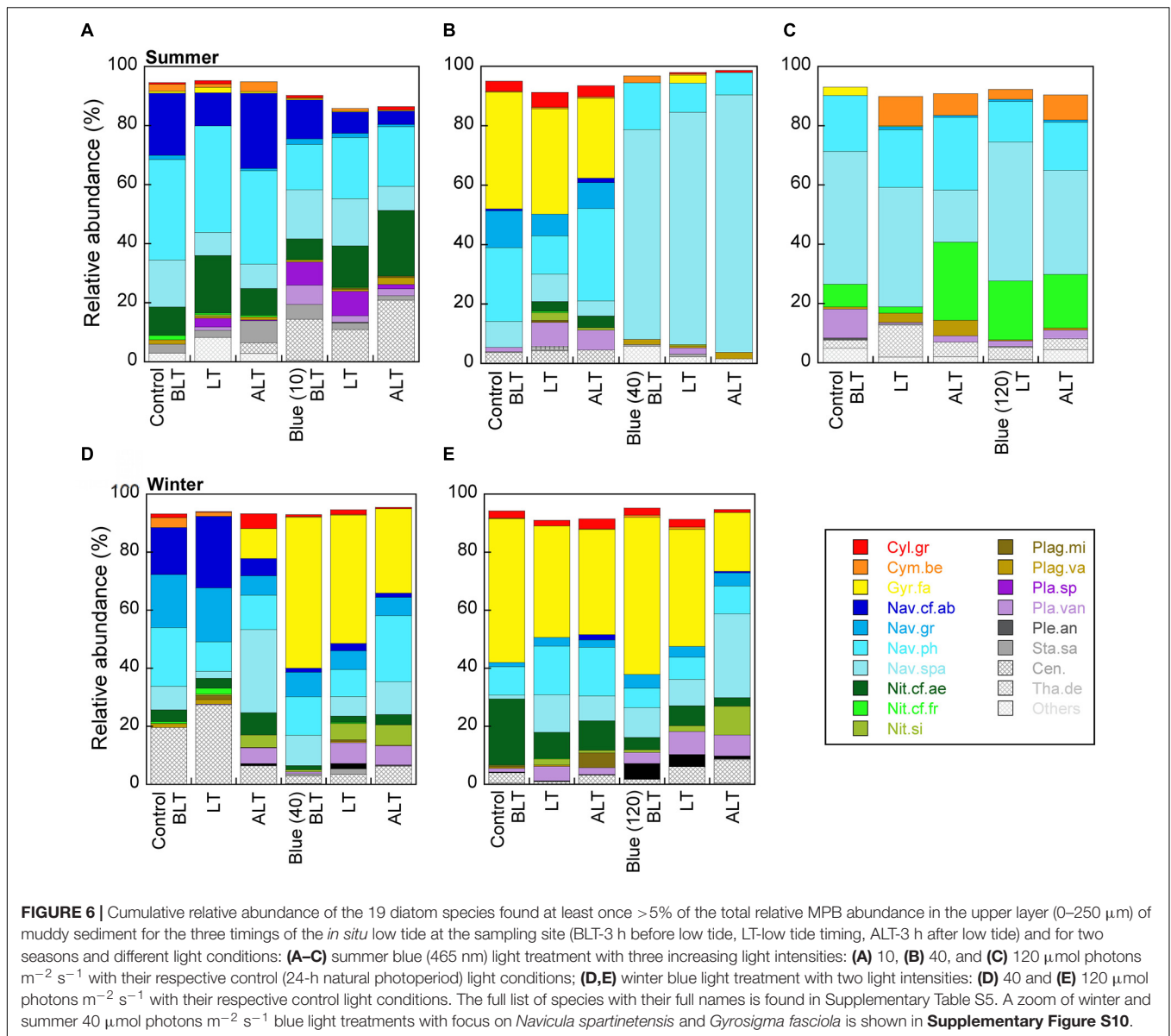
20%; (ii) for *N. spartinetensis*, a BL-10 and BL-120 did not induce changes in abundance relative to control light conditions (Figures 6A,C); the same was true for *G. fasciola* response to BL-120 (BL-10 was not tested). Finally, and to a much lower extent, other *Navicula* species (especially *Navicula phyllepta* and *Navicula gregaria*) might show a positive response to BL (Figure 6 and Supplementary Figure S9).

DISCUSSION

Monitoring of the Changes in the MPB PAB at the Surface of Sediment by Imaging-PAM Fluorimetry

The use of coupled Chl *a* fluorescence with imaging techniques has proven to be of paramount interest to monitor the temporal changes in MPB biomass, in particular with an Imaging-PAM fluorometer (Du et al., 2010; Vieira et al., 2013; Blommaert et al., 2018). Imaging of Chl *a* fluorescence allows assessment of the dynamics of the “photosynthetically active (or productive) biomass” (PAB) at the surface of sediment (Herlory et al., 2005; Guarini et al., 2008). In the present study, we have used Imaging-PAM fluorimetry to assess changes in PAB in a continuous mode with measurements every 900 s over several days with less than 15-min stop for watering and mixing the sediment tray every 24 h. The main advantage of imaging-fluorimetry compared to other fluorimeters (for instance, the Water-PAM fiber version or the Diving-PAM) is its non-invasive feature; there is no contact with the sediment surface (Vieira et al., 2013). This way, (i) MPB biofilm structure is not disturbed, (ii) its light microenvironment is less biased, and (iii) changes in MPB abundance can be followed

simultaneously with regular measurement of photosynthetic performances (for instance, here, ΦPSII_{inst}) taking into account potential limitations (Vieira et al., 2013). More precisely here, we have monitored F_t , the “instantaneous” Chl *a* fluorescence yield as measured with the detecting beam of the Imaging-PAM. The interest in using F_t instead of the well-known MPB biomass proxy F_0 (Consalvey et al., 2005) was to avoid darkening of the MPB, which is known to induce counteracting downward migration, and would have therefore disrupted rhythmic dynamics in MPB migration. Stock et al. (2019) also noticed that F_0 could be successfully replaced by F_t under some experimental conditions [see also Ni Longphuiert et al. (2006)]. We verified that F_t was well correlated to the Chl *a* content of sediment surface ($R^2 = 0.84$), a correlation as strong as the ones previously reported between F_t (Ni Longphuiert et al., 2006) or F_0 (Honeywill et al., 2002; Serôdio et al., 2006) and Chl *a*. Although the detecting beam was used with minimal irradiance settings, to be as close as possible to non-actinic conditions, still $0.81 \pm 0.08 \mu\text{mol photons m}^{-2} \text{s}^{-1}$ were generated at the surface of sediment trays. It appeared, *a posteriori*, that this intensity of blue wavelength (470 nm) was sufficiently effective in triggering positive, albeit limited, upward migration of MPB (see section “Influence of the Light Spectrum on the Amount of PAB Present at the Surface of Sediment: Specific Effect of BL Wavelengths”). This effect was most obvious with the $5.1\% \pm 4.0\%$ total PAB at night (as compared to control light conditions), which is not expected to occur (see section “What Influences the Most the Amount of PAB Present at the Surface of Sediment: Tides or Light?”). This is nevertheless consistent with earlier findings reporting an effect of irradiances as low as 0.5 to 1 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ in triggering the directional motility of epipellic diatoms (Paterson, 1986; Cohn and Weitzell, 1996; Wenderoth and Rhiel, 2004). There would be



two ways of avoiding this drawback: (1) instead of measuring F_t , to use the red and near-infrared LEDs of the Imaging-PAM LED panel to measure an NDVI index shown to be well correlated with Chl *a* sediment content (Blommaert et al., 2018); (2) to replace the blue Imaging-PAM measuring head by the red (650–660 nm) one for measuring F_t , which most likely (see section “Influence of the Light Spectrum on the Amount of PAB Present at the Surface of Sediment: Specific Effect of BL Wavelengths”) will not trigger MPB migration at night.

These minor technical limitations being considered, our experimental setup allowed us to reliably monitor MPB migration dynamics *ex situ*. The patterns we observed in muddy sediment dominated by epipellic diatoms at different times of the day and as a function of LT timings (i.e., at solar noon or close to sunset and/or sunrise, see section “Vertical Migration Pattern of MPB as a Function of Sediment Type, LT Timing, and

Seasons”) were well in accordance with those accurately described decades ago both *in situ* and in the laboratory (Consalvey et al., 2004). Of primary importance here was the checking of several prerequisites for further experimental manipulation. As others before (Consalvey et al., 2004), (i) we could observe a migratory rhythm of natural MPB assemblages in laboratory-controlled conditions and without any artificially recreated tidal rhythm (i.e., alternation of emersion/immersion); (ii) this migratory rhythm was precisely related to the *in situ* LT timings (i.e., corresponding to the lowest water level at the sampling site) and to the day length (photoperiod); (iii) migratory rhythm was maintained for 5 ± 1 consecutive days with only sea-watering and sediment mixing every 24 h at the exact timing of *in situ* HT; (iv) migratory rhythm was maintained whatever the season at which the sediment was sampled (albeit the amplitude was different, see below); (v) the photosynthetic potential of MBP

(Φ PSII $_{inst}$) was preserved. In fact, our conditioning of sediment was good enough, and our experimental setup sensitive enough so that we could record the MPB activity even in sandy sediment dominated by epipsammic diatoms. Epipsammic species are generally immotile except for representatives that are capable of micromovements within the sphere of the sediment particle to which they are attached, or from a particle to another (Harper, 1969; Round, 1979). Here, when compared to muddy sediment, we could observe (i) a stronger night basal Chl *a* fluorescence signal probably due to the deeper penetration of the Imaging-PAM detecting beam in the coarser sediment, and (ii) a slower, weaker but sustained increase in MPB abundance at the surface of sediment around daytime solar noon *in situ* LT timing that we interpret as the motile epipsammic and/or the small proportion of epipelagic diatoms moving toward the light source (Harper, 1969; Cartaxana et al., 2011), and thus increasing the fluorescence signal. By compiling all of our MPB migration profiles, we could draw a general scheme of surface PAB changes as a function of tidal amplitude, LT timing, and seasons, at our study site. Because the later was upper-shore (i.e., 500 m from the shore, approximately 4 m above the marine chart datum), MPB was often emerged, even for low tidal amplitude. This situation likely explains why we did not observe any clear correlation in PAB changes with the *in situ* tidal amplitude and LT timing (see also Brotas et al., 1995; Jesus et al., 2009; Coelho et al., 2011). However, surface PAB was strongly related to the sampling season with the highest values observed by the end of winter–beginning of spring. This general seasonal pattern strikingly matches the one recorded through the analysis of a 15-year satellite remote sensing time-series over the Brouage mudflat located just south of our sampling site (Méléder et al., unpublished). This result is of crucial importance for future up-scaling approaches for the estimation of intertidal mudflat primary production (Daggers et al., 2018), the present experimental setup providing a reliable intermediary step between *in situ* measurements and satellite images.

What Influences the Most the Amount of PAB Present at the Surface of Sediment: Tides or Light?

It is generally accepted that the rhythmic MPB vertical migration in intertidal sediment and the surface PAB accumulation are governed by the tidal and diurnal cycles together (Consalvey et al., 2004; Coelho et al., 2011). There are nevertheless situations where tides have a more limited influence and/or light a stronger influence; in such conditions, MPB migration strongly matches with diurnal photoperiodic and irradiance (see below) changes (see, for instance, Mitbavkar and Anil, 2004; Ni Longphuiert et al., 2006). It highlights the possibility to disentangle the tight link between the tidal and diurnal rhythmicity of MPB migration. We thus manipulated the 24-h natural photoperiod of MPB in order to try desynchronizing MPB migration pattern from the tidal and/or diurnal cycles. Differently from previous works (Kwon et al., 2012; Haro et al., 2019), we did not submerge the sediment in order to avoid potentially exacerbated photoperiodic response as in subtidal

sediments (Ni Longphuiert et al., 2006). Our experiments have shown that (i) MPB migration can persist for 5 ± 1 days *ex situ* following *in situ* tidal cycle and photoperiod even when permanently maintained emerged (although homogenized with seawater every 24 h); (ii) in neap tide situation (i.e., LT timings close to sunset and sunrise), MPB migration occurs irrespective of the photoperiod and is primarily modeled onto the tidal cycle; (iii) under 24-h darkness or light exposure, MPB migration goes on matching the LT timing of the subjective day or night; (iii) within few days, it is possible to completely desynchronize MPB migration from the tidal cycle to force it matching photoperiod such as in *in situ* subtidal sediments (Ni Longphuiert et al., 2006) or intertidal sediments immersed *ex situ* (Paterson, 1986; Haro et al., 2019). These observations confirm and complete previous ones [see the synthesis by Consalvey et al. (2004), as well as more recent works where MPB migration was experimentally manipulated *ex situ* (Coelho et al., 2011; Kwon et al., 2012; Haro et al., 2019)]. Also important are the many *in situ* observations that (see Happey-Wood and Jones, 1988; Consalvey et al., 2004; Coelho et al., 2011) (i) MPB at the surface of sediment appears to “anticipate” the incoming tide by migrating downward, although this behavior is not systematic; (ii) MPB migrates downward even during emersion at dusk or when darkened.

Taken all together, these features support the endogenous nature of intertidal MPB migration rhythmicity, which is first modeled onto the tidal cycle [so-called “lunidian clock” phased on the lunar day length and cycle (Palmer, 2000)], which can be modulated (i.e., entrained and/or reset; Haro et al., 2019), potentiated, and even overridden, by the diurnal cycle, also potentially controlled by an endogenous circadian clock, in some situations. Quantification of the “potentiating” effect of non-stressful light can be attempted using PAB values under prolonged darkness and continuous light exposures: respectively, $39\% \pm 2\%$ and $144\% \pm 3\%$ of control light conditions, strikingly similar to previous reports (approximately 40% in darkness and approximately 130–155% at optimal irradiance) (Sauer et al., 2002; Coelho et al., 2011). These values would mean that light can potentiate the tidal effect on surface PAB accumulation by a factor of 1.6 under 24-h natural photoperiod conditions to more than 2.5 in optimal laboratory conditions (see also Coelho et al., 2011). It was proposed before that endogenous clock(s) would control cell gravitaxy to match the tidal cycle and cell phototaxy to match the diurnal cycle (see Mitbavkar and Anil, 2004; Coelho et al., 2011; Frankenbach et al., 2014). Saburova and Polikarpov (2003) additionally proposed that the central biological driver of rhythmic MPB migration is the cell division cycle and its spatial disconnection from photosynthesis; that is, the cell cycle occurs in the stable and nutrient-rich deep sediment layers during immersion/at night, and it needs the photochemical energy and organic carbon that is generated and stored at the surface of sediment during daytime emersion (Underwood et al., 2005; Marques Da Silva et al., 2017). This temporal (light vs. dark periods) metabolic decoupling in diatoms has been further demonstrated (Skogen Chauton et al., 2013). The

endogenously controlled rhythmic MPB migration, modeled onto intertidal flat environmental conditions, would allow MPB to perform surface and depth biological processes in the most optimal conditions. Although endogenous circadian clocks, their components and regulation, have been well detailed in several algal lineages, they remain uncharacterized in diatoms (Noordally and Millar, 2015), with the exception of the recent discovery of a time-keeping molecular component, which controls the diel rhythmic expression of many genes (Annunziata et al., 2019).

Both lunar and solar day major drivers can be in turn modulated by other environmental stimuli (temperature, nutrients, salinity, pH, etc.; see Consalvey et al., 2004), and by other light features: light spectrum (see section “Influence of the Light Spectrum on the Amount of PAB Present at the Surface of Sediment: Specific Effect of BL Wavelengths”) and irradiance. Indeed, during emersion, the light availability at the surface and in sediment is also defined by the course of the sun during the daytime period, by open-field-like incident irradiance fluctuations and by the sediment-dependent light penetration, which varies with several features [particle size, content in pore water, etc. (Kuhl and Jorgensen, 1994)]. Many studies have reported how the level of irradiance controls epipellic diatom phototaxis in a biphasic way. While up to a certain irradiance threshold light promotes upward migration and the formation of the MPB biofilm, beyond this threshold it conversely induces downward migration and decreases surface PAB (see, for instance, Serôdio et al., 2006; Laviale et al., 2016). It is believed that motility along the sediment light gradient allows MPB both to optimize photosynthesis and to avoid stressful light conditions (i.e., above the photosynthetic capacity of cells). In the present study, the phototactic behavior of epipellic MPB was well illustrated by the PAB present at the surface of sediment as a function of “white” light irradiance with a maximum reached for 120 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$. Similar values (i.e., range 100–250 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$) were also reported in previous studies on both MPB natural communities (Perkins et al., 2002; Serôdio et al., 2006; Laviale et al., 2016; Du et al., 2018) and isolated epipellic diatom species (McLachlan et al., 2009; Du et al., 2010; Ezequiel et al., 2015). This feature was essential for two reasons: (i) it allowed us to validate the relevance of our experimental setup, and especially its ability to well preserve MPB light response in the sediment, and (ii) it allowed us to draw reference light conditions for maximizing the surface PAB, two necessary conditions for further testing together the effect of light intensity and spectrum on MPB migration.

Influence of the Light Spectrum on the Amount of PAB Present at the Surface of Sediment: Specific Effect of BL Wavelengths

In the present study, surface PAB accumulation was positively influenced by blue (465 nm) and possibly by green (530 nm) wavelengths, but not by red ones (677 nm, with or without far red). Although green light generated a similar PAB

accumulation than BL for the same intensity (40 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$), our experimental setup did not allow to test higher green light intensities to unambiguously conclude the responses to green light. It was previously demonstrated that green light (>510–520 nm) generally has a lower effect than BL (450 \pm 20 nm) on phototactic directional cell movements (Nultsch, 1971; Cohn et al., 2004, 2015, 2016). Differently, red light of low or moderate intensity usually has no/little effect or negative effect on directional motility of diatoms and their accumulation, although this response is species-specific (Cohn and Weitzell, 1996; McLachlan et al., 2009; Cohn et al., 2015). These reports partially explain the action spectrum of MPB abundance at the surface of sediment at low irradiance (5 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$) showing a strong positive effect of blue wavelengths (440 \pm 10 nm), limited effect of greens (500–550 nm), and a moderate effect of red (680 nm) (Wenderoth and Rhiel, 2004).

Here, as regards to the specific effect of blue wavelengths, we have found that (i) BL induced a $\times 1.8 \pm 0.4$ total PAB increase compared to the “white” light control at the same intensity of 120 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$; (ii) PAB accumulation was irradiance-dependent with a positive effect from 10 to 120 $\mu\text{mol BL photons m}^{-2} \text{s}^{-1}$; (iii) PAB accumulation was species-specific with two taxa, namely, *N. spartinetensis* and *G. fasciola* (and probably other *Navicula* species to a lesser extent), which strongly reacted (i.e., from 0 to 10% relative presence at the surface of sediment to up to more than 80%) at 40 $\mu\text{mol BL photons m}^{-2} \text{s}^{-1}$; (iv) their surface accumulation was independent of the season: the BL effect was observed with both winter and summer sediments for a particular species. These features fit well with previous reports. In general, blue wavelengths have a positive effect (up to a certain irradiance threshold, see Cohn et al., 2004; McLachlan et al., 2009) on surface PAB ($\times 1.4$ to $\times 1.8$, Wenderoth and Rhiel, 2004) by modulating the speed to which epipellic diatoms move directionally (Nultsch, 1971; Cohn and Weitzell, 1996; McLachlan et al., 2009; Cohn et al., 2015). This BL photosensitivity is species-specific (McLachlan et al., 2009; Cohn et al., 2015), and it is detected at least at the tips of pennate cells (Cohn et al., 1999). The way blue wavelengths act at the cellular level and how the “BL signal” might be transduced is still unclear in diatoms, although significant recent advances have been made (Jaubert et al., 2017). It was early proposed (Nultsch, 1971) that pigments, chlorophylls and carotenoids, should act as primary harvesters and transducers to trigger the previously described action spectrum of MPB motility, and especially photokinesis (i.e., change of motility speed; Wenderoth and Rhiel, 2004). The latter would directly depend on the energy provided by photosynthesis, which itself is driven by actinic wavelengths, and obviously BL. In that respect, the optical properties of the photonic crystal-like structures of the diatom frustule are highly relevant; they have been shown to shape the availability of light and of specific wavelengths inside the cell, thereby potentiating pigment light-harvesting and photosynthetic efficiency (Goessling et al., 2018a,b). Interestingly, the frustule of one of our most BL-reactive species, *G. fasciola*,

was shown to be transparent when the frustule of other epipellic species significantly (up to 20%) attenuated the transmission of blue wavelengths (Goessling et al., 2018a). While BL attenuation can be advantageous to decrease potentially harmful incident irradiance, its nearly 100% transmission under low/moderate irradiance could support specific BL signaling inside the cell. These interspecies differences in frustule optical properties were claimed to additionally modulate the light availability inside the biofilm itself (Goessling et al., 2018a). Taken together with the fact that a species response to BL can be modulated by the presence of other species in its close surrounding (Cohn et al., 2016), it points out to the importance of the species consortium composition, which varies year around (this study, Haubojs et al., 2005). Therefore, wavelengths, together with irradiance, would fine tune the differential rhythmic MPB migratory behavior and surface PAB accumulation versus the sediment light climate at the cell, species, and consortium (i.e., biofilm) levels.

However, in the present study, upon 24-h exposure, we did not observe any specific BL effect on (i) MPB pigment content, and especially carotenoids, independent of the irradiance, of the season, and of the sediment depth (down to 600 μm); (ii) MPB photosynthetic performances, independent of the irradiance. Therefore, the positive BL effect we observed was based on molecular cell components that were already present at the beginning of the light treatment (such as pigments), and it was not due to a surge in photosynthetic performance, that is, higher surface PAB accumulating because of more diatom cells photosynthesizing better under BL. Although we do not question the importance of the frustule optical properties and of photopigments in triggering the BL-enhanced PAB with the irradiance-dependent PAB increase being a proof of such role, we aim at seeking for alternative complementary answers. Photoreceptors have been proposed before to play a central role in the positive upward phototaxis of epipellic MPB to blue wavelengths (Wenderoth and Rhiel, 2004; McLachlan et al., 2009). In the recent years, three types of BL photoreceptors have been identified in planktonic diatom models including cryptochromes and a family unique to stramenopiles, aureochromes (Jaubert et al., 2017). Aureochromes control the onset of diatom cell division (mitosis) (Huysman et al., 2013), and BL stimulates growth rate and biofilm formation of the epipellic representative *Navicula* species (Cao et al., 2013). Therefore, the BL increase in surface PAB we observed could also be due to the proliferation of diatom cells. Because blue wavelengths are rapidly attenuated with sediment depth (Kuhl and Jorgensen, 1994), it is possible that BL, in conjunction with the necessary level of photochemical energy and organic carbon stored during emersion (Underwood et al., 2005; Marques Da Silva et al., 2017), could act as a trigger for mitosis when migrating downward, which, according to Saburova and Polikarpov (2003), are coupled. Furthermore, recent highlights on the role of photoreceptors in the rhythmic light response of diatoms (see Jaubert et al., 2017 for a synthesis) point out to a highly complex regulatory molecular network of light-dependent (such as the photoperiod) and independent (such

as the tidal cycle) biological rhythms. It is very likely that sophisticated cross-talks between light perception (by frustule, pigments, and photoreceptors), photosynthetic metabolism, and endogenous regulators (circadian clock components) occur to finely tune the light response of diatoms beyond photoperiod (see section “What Influences the Most the Amount of PAB Present at the Surface of Sediment: Tides or Light?”). Obviously, such a network would be a strong asset for epipellic diatoms in coping with the multifaceted intertidal sediment light climate.

CONCLUSION

The present study strengthens the view that the vertical migratory rhythm of motile MPB diatoms in intertidal sediment is of endogenous nature and is first mapped onto the tidal cycle with the migration rhythm driven by a “lunidian clock” phased on the lunar day length and cycle. Migration rhythmicity is further modulated, and even overridden in some conditions, by light, including the diurnal cycle and the irradiance level during daytime periods. Optimal light conditions can significantly “potentiate” the tidal effect on the accumulation of PAB at the surface of sediment during emersion. Additionally, we found a strong positive effect of blue wavelengths (465 nm) on surface PAB accumulation as compared to others (white, green, and red lights). This effect is intensity- and species-dependent (i.e., some species strongly react to BL, whereas others do not). We propose that, in addition to photopigments and frustule optical properties, BL photoreceptors (cryptochromes and aureochromes) might play a crucial role in regulating the presence and abundance of PAB at the surface of intertidal sediment. We found two new potential model species, *N. spartinetensis* and *G. fasciola*, which strongly react to blue wavelengths. The fact that *N. spartinetensis* shows a much smaller size than *G. fasciola*, and probably a different migratory behavior (see Wenderoth et al., 2004; Underwood et al., 2005) is an additional asset. The complementary use of the epipellic model *Seminavis robusta* (Blommaert et al., 2017, 2018), which genome has been sequenced, will allow to search for molecular players, such as timekeepers (Annunziata et al., 2019), photoreceptors (Jaubert et al., 2017), and diurnal-responsive genes (see Meyer et al., 2003; Blommaert et al., 2020), of the rhythmic migratory light-tuned response of intertidal epipellic MPB.

DATA AVAILABILITY STATEMENT

All datasets generated for this study are included in the article/**Supplementary Material**.

AUTHOR CONTRIBUTIONS

AB and JL contributed to the conception and design of the study. AB performed the experiments. AB, VM, and

JL performed samples and data analyses. AB wrote the first draft of the manuscript. All authors contributed to manuscript revision, read and approved the submitted version.

FUNDING

This work was supported by the Centre National de la Recherche Scientifique-CNRS, the University of La Rochelle-ULR, the Contrat Plant Etat Région-CPER "Littoral" 2007–2013, the Region Poitou-Charentes-CG17, and by the Natural Sciences and Engineering Research Council-NSERC (Discovery grant to JL).

REFERENCES

- Admiraal, W. (1984). "The ecology of estuarine sediment inhabiting diatoms," in *Progress Phycology Research*, eds F. E. Round and D. J. Chapman (Bristol: Biopress Limited), 269–314.
- Annuziata, R., Rittera, A., Fortunato, A. E., Manzotti, A., Cheminant-Navarro, S., Agier, N., et al. (2019). bHLH-PAS protein RITMO1 regulates diel biological rhythms in the marine diatom *Phaeodactylum tricornerutum*. *Proc. Natl. Acad. Sci. U.S.A.* 116, 13137–13142. doi: 10.1073/pnas.1819660116
- Apoya-Horton, M. D., Yin, L., Underwood, G. J. C., and Gretz, M. R. (2006). Movement modalities and responses to environmental changes of the mudflat diatom *Cylindrotheca closterium* (Bacillariophyceae). *J. Phycol.* 42, 379–390. doi: 10.1111/j.1529-8817.2006.00194.x
- Barnett, A., Méléder, V., Blommaert, L., Lepetit, B., Gaudin, P., Vyverman, W., et al. (2015). Growth form defines physiological photoprotective capacity in intertidal benthic diatoms. *ISME J.* 9, 32–45. doi: 10.1038/ismej.2014.105
- Bertrand, J. (2008). Mouvements des diatomées VIII: synthèse et hypothèse. *Diatom Res.* 23, 19–29. doi: 10.1080/0269249x.2008.9705734
- Blanchard, G., Guarini, J.-M., Gros, P., and Richard, P. (1997). Seasonal effect on the relationship between photosynthetic capacity of intertidal microphytobenthos and temperature. *J. Phycol.* 33, 723–728. doi: 10.1111/j.0022-3646.1997.00723.x
- Blommaert, L., Huysman, M. J., Vyverman, W., Lavaud, J., and Sabbe, K. (2017). Contrasting NPQ dynamics and xanthophyll cycling in a motile and a non-motile intertidal benthic diatom. *Limnol. Oceanogr.* 62, 1466–1479. doi: 10.1002/lno.10511
- Blommaert, L., Lavaud, J., Vyverman, W., and Sabbe, K. (2018). Behavioural versus physiological photoprotection in epipellic and epipsammic benthic diatoms. *Eur. J. Phycol.* 53, 146–155. doi: 10.1080/09670262.2017.1397197
- Blommaert, L., Vancaester, E., Huysman, M. J. J., Osuna-Cruz, C. M., D'hondt, S., Lavaud, J., et al. (2020). Light regulation of LHCX genes in the benthic diatom *Seminavis robusta*. *Front. Mar. Sci.* 7:192. doi: 10.3389/fmars.2020.00192
- Brotas, V., Cabrita, T., Portugal, A., Seródio, J., and Catarino, F. (1995). Spatio-temporal distribution of the microphytobenthic biomass in intertidal flats of Tagus Estuary (Portugal). *Hydrobiologia* 300, 93–104. doi: 10.1007/bf00024451
- Cao, S., Wang, J., and Chen, D. (2013). Settlement and cell division of diatom *Navicula* can be influenced by light of various qualities and intensities. *J. Basic Microbiol.* 53, 884–894. doi: 10.1002/jobm.201200315
- Cartaxana, P., Cruz, S., Gameiro, C., and Kühl, M. (2016a). Regulation of intertidal microphytobenthos photosynthesis over a diel emersion period is strongly affected by diatom migration patterns. *Front. Microbiol.* 7:872. doi: 10.3389/fmicb.2016.00872
- Cartaxana, P., Ribeiro, L., Goessling, J. W., Cruz, S., and Kühl, M. (2016b). Light and O₂ microenvironments in two contrasting diatom-dominated coastal sediments. *Mar. Ecol. Progr. Ser.* 545, 35–47. doi: 10.3354/meps11630
- Cartaxana, P., Ruivo, M., Hubas, C., Davidson, I., Seródio, J., and Jesus, B. (2011). Physiological versus behavioral photoprotection in intertidal epipellic and epipsammic benthic diatom communities. *J. Exp. Mar. Biol. Ecol.* 405, 120–127. doi: 10.1016/j.jembe.2011.05.027

ACKNOWLEDGMENTS

The authors thank I. Benyoucef for his help with the preparation of definitive slides, the identification and the counting of diatom cells, J.-L. Mouget for the access to the Water-PAM, X. Cousin for the access to the cryomicrotome, and D. Campbell for the critical reading of the manuscript.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fmars.2020.00212/full#supplementary-material>

- Coelho, H., Vieira, S., and Seródio, J. (2011). Endogenous versus environmental control of vertical migration by intertidal benthic microalgae. *Eur. J. Phycol.* 46, 271–281. doi: 10.1080/09670262.2011.598242
- Cohn, S. A. (2001). "Photo-stimulated effects on diatom motility," in *Photomovement*, eds D.-P. Häder and M. Lebert (Amsterdam: Elsevier), 375–401. doi: 10.1016/s1568-461x(01)80017-x
- Cohn, S. A., Bahena, M., Davis, J. T., Ragland, R. L., Rauschenberg, C. D., and Smith, B. J. (2004). Characterisation of the diatom photophobic response to high irradiance. *Diatom Res.* 19, 167–179. doi: 10.1080/0269249x.2004.9705869
- Cohn, S. A., Dunbar, S., Ragland, R. L., Schulze, J., Suchar, A., Weiss, J., et al. (2016). Analysis of light quality and assemblage composition on diatom motility and accumulation rate. *Diatom Res.* 31, 173–184. doi: 10.1080/0269249x.2016.1193058
- Cohn, S. A., Halpin, D., Hawley, N., Ismail, A., Kaplan, Z., Kordes, T., et al. (2015). Comparative analysis of light-stimulated motility responses in three diatom species. *Diatom Res.* 30, 213–225. doi: 10.1080/0269249x.2015.1058295
- Cohn, S. A., Spurck, T. P., and Pickett-Heaps, J. D. (1999). High energy irradiation at the leading tip of moving diatoms causes a rapid change of cell direction. *Diatom Res.* 14, 193–206. doi: 10.1080/0269249x.1999.9705466
- Cohn, S. A., and Weitzell, R. E. (1996). Ecological considerations of diatom cell motility I. Characterization of motility and adhesion in four diatom species. *J. Phycol.* 32, 928–939. doi: 10.1111/j.0022-3646.1996.00928.x
- Consalvey, M., Paterson, D. M., and Underwood, G. J. C. (2004). The ups and downs of life in a benthic biofilm: migration of benthic diatoms. *Diatom Res.* 19, 181–202. doi: 10.1080/0269249x.2004.9705870
- Consalvey, M., Perkins, G. R., Paterson, D. M., and Underwood, G. J. C. (2005). PAM fluorescence: a beginners guide for benthic diatomists. *Diatom Res.* 20, 1–22. doi: 10.1080/0269249x.2005.9705619
- Daggers, T. D., Kromkamp, J. C., Herm, P. M. J., and Van Der Wal, D. (2018). A model to assess microphytobenthic primary production in tidal systems using satellite remote sensing. *Remote Sens. Environ.* 211, 129–145. doi: 10.1016/j.rse.2018.03.037
- Du, G. Y., Oak, J. H., Li, H., and Chung, I. K. (2010). Effect of light and sediment grain size on the vertical migration of benthic diatoms. *Algae* 25, 133–140. doi: 10.4490/algae.2010.25.3.133
- Du, G. Y., Yan, H., Liu, C., and Moa, Y. (2018). Behavioral and physiological photoresponses to light intensity by intertidal microphytobenthos. *Chinese J. Oceanol. Limnol.* 36, 293–304. doi: 10.1007/s00343-017-6099-0
- Eilers, P. H. C., and Peeters, J. C. H. (1988). A model for the relationship between light intensity and the rate of photosynthesis in phytoplankton. *Ecol. Model.* 42, 199–215. doi: 10.1016/0304-3800(88)90057-9
- Ezequiel, J., Laviale, M., Frankenbach, S., Cartaxana, P., and Seródio, J. (2015). Photoacclimation state determines the photobehaviour of motile microalgae: the case of a benthic diatom. *J. Exp. Mar. Biol. Ecol.* 468, 11–20. doi: 10.1016/j.jembe.2015.03.004
- Frankenbach, S., Pais, C., Martinez, M., Laviale, M., Ezequiel, J., and Seródio, J. (2014). Evidence for gravitactic behaviour in benthic diatoms. *Eur. J. Phycol.* 49, 429–435. doi: 10.1080/09670262.2014.974218

- Goessling, J. W., Frankenbach, S., Ribeiro, L., Seródio, J., and Kühl, M. (2018a). Modulation of the light field related to valve optical properties of raphid diatoms: implications for niche differentiation in the microphytobenthos. *Mar. Ecol. Progr. Ser.* 588, 29–42. doi: 10.3354/meps12456
- Goessling, J. W., Yanyan, Y., Cartaxana, P., Maibohm, C., Rickelt, L. F., Trampe, E. C. L., et al. (2018b). Structure-based optics of centric diatom frustules: modulation of the *in vivo* light field for efficient diatom photosynthesis. *New Phytol.* 219, 122–134. doi: 10.1111/nph.15149
- Guarini, J.-M., Blanchard, G., and Richard, P. (2006). “Modelling the dynamics of the microphytobenthic biomass and primary production in European intertidal mudflats,” in *Functioning of Microphytobenthos in Estuaries*, eds J. Kromkamp, J. F. C. De Brouwer, G. Blanchard, R. M. Forster, and V. Créach (Amsterdam: Royal Netherlands Academy of Arts and Sciences), 187–226.
- Guarini, J.-M., Saric, N., and Moritz, C. (2008). Modelling the dynamics of the microalgal biomass in semi-enclosed shallow-water ecosystems. *Ecol. Model.* 211, 267–278. doi: 10.1016/j.ecolmodel.2007.09.011
- Happey-Wood, C. M., and Jones, P. (1988). Rhythms of vertical migration and motility in intertidal benthic diatoms with particular reference to *Pleurosigma angulatum*. *Diatom Res.* 3, 83–93. doi: 10.1080/0269249x.1988.9705018
- Haro, S., Bohórquez, J., Lara, M., Garcia-Robledo, E., González, C. J., Crespo, J. M., et al. (2019). Diel patterns of microphytobenthic primary production in intertidal sediments: the role of photoperiod on the vertical migration circadian rhythm. *Sci. Rep.* 9:13376.
- Harper, M. A. (1969). Movement and migration of diatoms on sand grains. *Br. Phycol. J.* 4, 97–103. doi: 10.1080/00071616900650081
- Haubois, A.-G., Sylvestre, F., Guarini, J.-M., Richard, P., and Blanchard, G. F. (2005). Spatio-temporal structure of the epipellic diatom assemblage from an intertidal mudflat in Marennes-Oléron Bay, France. *Estuar. Coast Shelf Sci.* 64, 385–394. doi: 10.1016/j.ecss.2005.03.004
- Herlory, O., Blanchard, G. F., Planche, S., Huet, V., and Richard, P. (2005). Does the size of the microphytobenthic biofilm on intertidal mudflats depend on the available photosynthetic biomass? *Mar. Ecol. Progr. Ser.* 298, 95–100. doi: 10.3354/meps298095
- Herlory, O., Guarini, J.-M., Richard, P., and Blanchard, G. F. (2004). Microstructure of microphytobenthic biofilm and its spatio-temporal dynamics in an intertidal mudflat (Aiguillon Bay, France). *Mar. Ecol. Progr. Ser.* 282, 33–44. doi: 10.3354/meps282033
- Honeywill, C., Paterson, D. M., and Hagerthey, S. E. (2002). Determination of microphytobenthic biomass using pulse-amplitude modulated minimum fluorescence. *Eur. J. Phycol.* 37, 485–492. doi: 10.1017/s0967026202003888
- Huysman, M. J., Fortunato, A. E., Matthijs, M., Costa, B. S., Vanderhaeghen, R., Van Den Daele, H., et al. (2013). AUREOCHROME1a-mediated induction of the diatom-specific cyclin dsCYC2 controls the onset of cell division in diatoms (*Phaeodactylum tricornutum*). *Plant Cell* 25, 215–228. doi: 10.1105/tpc.112.106377
- Jakob, T., Goss, R., and Wilhelm, C. (1999). Activation of diadinoxanthin de-epoxidase due to a chlororespiratory proton gradient in the dark in the diatom *Phaeodactylum tricornutum*. *Plant Biol.* 1, 76–82. doi: 10.1111/j.1438-8677.1999.tb00711.x
- Jaubert, M., Bouly, J.-P., D'alcala, M. R., and Falcitore, A. (2017). Light sensing and responses in marine microalgae. *Trends Plant Biol.* 37, 70–77. doi: 10.1016/j.pbi.2017.03.005
- Jesus, B., Brotas, V., Ribeiro, L., Mendes, C. R., Cartaxana, P., and Paterson, D. M. (2009). Adaptations of microphytobenthos assemblages to sediment type and tidal position. *Cont. Shelf Res.* 29, 1624–1634. doi: 10.1016/j.csr.2009.05.006
- Kooistra, W. H. C. F., Gersonde, R. K., Medlin, L. G., and Mann, D. (2007). “The origin and evolution of the diatoms: their adaptation to a planktonic existence,” in *Evolution of Primary Producers in the Sea*, eds P. G. Falkowski and A. H. Knoll (Burlington: Academic Press), 207–249. doi: 10.1016/b978-012370518-1/50012-6
- Kuhl, M., and Jorgensen, B. B. (1994). The light field of microbenthic communities: radiance distribution and microscale optics of sandy coastal sediments. *Limnol. Oceanogr.* 39, 1368–1398. doi: 10.4319/lo.1994.39.6.1368
- Kwon, B.-O., Khim, J. S., Park, J., Ryu, J., Kang, S.-G., and Koh, C.-H. (2012). Short-term variability of microphytobenthic primary production associated with *in situ* diel and tidal conditions. *Estuar. Coast Shelf Sci.* 112, 236–242. doi: 10.1016/j.ecss.2012.07.029
- Laviale, M., Ezequiel, J., Pais, C., Cartaxana, P., and Seródio, J. (2015). The “crème brûlée” sampler: a new high-resolution method for the fast vertical sampling of intertidal fine sediments. *J. Exp. Mar. Biol. Ecol.* 468, 37–44. doi: 10.1016/j.jembe.2015.03.013
- Laviale, M., Frankenbach, S., and Seródio, J. (2016). The importance of being fast: comparative kinetics of vertical migration and non-photochemical quenching of benthic diatoms under light stress. *Mar. Biol.* 160:10.
- Lebreton, B., Rivaud, A., Picot, L., Prévost, B., Barillé, L., Sauzeau, T., et al. (2019). From ecological relevance of the ecosystem services concept to its sociopolitical use. The case study of intertidal bare mudflats in the Marennes-Oléron Bay, France. *Ocean Coast. Manage* 172, 41–54. doi: 10.1016/j.ocecoaman.2019.01.024
- Lefebvre, S., Mouget, J.-L., and Lavaud, J. (2011). Duration of rapid light curves for determining the photosynthetic activity of microphytobenthos biofilm *in situ*. *Aquat. Bot.* 95, 1–8. doi: 10.1016/j.aquabot.2011.02.010
- Lepetit, B., Sturm, S., Rogato, A., Gruber, A., Sachse, M., Falcitore, A., et al. (2013). High light acclimation in the secondary plastids containing diatom *Phaeodactylum tricornutum* is triggered by the redox state of the plastoquinone pool. *Plant Physiol.* 161, 853–865. doi: 10.1104/pp.112.207811
- Marques Da Silva, J., Cruz, S., and Cartaxana, P. (2017). Inorganic carbon availability in benthic diatom communities: photosynthesis and migration. *Phil. Trans. R. Soc. B* 372:20160398. doi: 10.1098/rstb.2016.0398
- McLachlan, D. H., Brownlee, C., Taylor, A. R., Geider, R. J., and Underwood, G. J. C. (2009). Light-induced motile responses of the estuarine benthic diatoms *Navicula Perminuta* and *Cylindrotheca Closterium* (Bacillariophyceae). *J. Phycol.* 45, 592–599. doi: 10.1111/j.1529-8817.2009.00681.x
- Méléder, V., Rincé, Y., Barillé, L., Gaudin, P., and Rosa, P. (2007). Spatiotemporal changes in microphytobenthos assemblages in a macrotidal flat (Bourgneuf Bay, France). *J. Phycol.* 43, 1177–1190. doi: 10.1111/j.1529-8817.2007.00423.x
- Meyer, T., Hust, M., Marquardt, J., Krumbein, W. E., and Rhiel, E. (2003). A methodological approach to investigate steady state fucoxanthin chlorophyll a/c binding protein mRNA levels in Wadden Sea sediments. *Internatl. Microbiol.* 6, 33–39. doi: 10.1007/s10123-003-0098-z
- Mitbavkar, S., and Anil, A. C. (2004). Vertical migratory rhythms of benthic diatoms in a tropical intertidal sand flat: influence of irradiance and tides. *Mar. Biol.* 145, 9–20.
- Nakov, T., Beaulieu, J. M., and Alverson, A. J. (2018). Accelerated diversification is related to life history and locomotion in a hyperdiverse lineage of microbial eukaryotes (Diatoms, Bacillariophyta). *New Phytol.* 219, 462–473. doi: 10.1111/nph.15137
- Ni Longphuiert, S., Leynaert, A., Guarini, J.-M., Chauvaud, L., Claquin, P., Herlory, O., et al. (2006). Discovery of microphytobenthos migration in the subtidal zone. *Mar. Ecol. Progr. Ser.* 328, 143–154. doi: 10.3354/meps328143
- Noordally, Z. B., and Millar, A. J. (2015). Clocks in algae. *Biochemistry* 54, 171–183. doi: 10.1021/bi501089x
- Nultsch, W. (1971). Phototactic and photokinetic action spectra of the diatom *Nitzschia communis*. *Photochem. Photobiol.* 14, 705–712. doi: 10.1111/j.1751-1097.1971.tb06209.x
- Palmer, J. D. (2000). The clocks controlling the tide-associated rhythms of intertidal animals. *BioEssays* 22, 32–37. doi: 10.1002/(sici)1521-1878(200001)22:1<32::aid-bies7>3.0.co;2-u
- Pante, E., and Simon-Bouhet, B. (2013). Marmap: a package for importing, plotting and analyzing bathymetric and topographic data in R. *PLoS One* 8:e73051. doi: 10.1371/journal.pone.0073051
- Paterson, D. M. (1986). The migratory behaviour of diatom assemblages in a laboratory tidal micro-ecosystem examined by low temperature scanning electron microscopy. *Diatom Res.* 1, 227–239. doi: 10.1080/0269249x.1986.9704971
- Paterson, D. M., and Hagerthey, S. E. (2001). “Microphytobenthos in contrasting coastal ecosystems: biology and dynamics,” in *Ecological Comparisons of Sedimentary Shores*, ed. K. Reise (Berlin: Springer-Verlag), 106–125.
- Perkins, R. G., Oxborough, K., Hanlon, A. R. M., Underwood, G. J. C., and Baker, N. R. (2002). Can chlorophyll fluorescence be used to estimate the rate of photosynthetic electron transport within microphytobenthic biofilms? *Mar. Ecol. Progr. Ser.* 228, 47–56. doi: 10.3354/meps228047

- Ribeiro, L. (2010). *Intertidal Benthic Diatoms of the Tagus Estuary: Taxonomic Composition and Spatial-Temporal Variation*. Ph.D. thesis, Universidade de Lisboa, Lisboa.
- Ribeiro, L., Brotas, V., Rince, Y., and Jesus, B. (2013). Structure and diversity of intertidal benthic diatom assemblages in contrasting shores: a case study from the Tagus estuary. *J. Phycol.* 49, 258–270. doi: 10.1111/jpy.12031
- Round, F. E. (1979). A diatom assemblage living below the surface of intertidal sand flats. *Mar. Biol.* 54, 219–223. doi: 10.1007/bf00395784
- Saburova, M. A., and Polikarpov, I. G. (2003). Diatom activity within soft sediments: behavioural and physiological processes. *Mar. Ecol. Progr. Ser.* 251, 115–126. doi: 10.3354/meps251115
- Sauer, J., Wenderoth, K., Maier, U. G., and Rhiel, E. (2002). Effects of salinity, light and time on the vertical migration of diatom assemblages. *Diatom Res.* 17, 189–203. doi: 10.1080/0269249x.2002.9705538
- Savelli, R., Dupuy, C., Barillé, L., Lerouxel, A., Guizien, K., Philippe, A., et al. (2018). On biotic and abiotic drivers of the microphytobenthos seasonal cycle in a temperate intertidal mudflat: a modelling study. *Biogeosciences* 15, 7243–7271. doi: 10.5194/bg-15-7243-2018
- Serôdio, J., Coelho, H., Vieira, S., and Cruz, S. (2006). Microphytobenthos vertical migratory photoresponse as characterised by light-response curves of surface biomass. *Estuar. Coast Shelf Sci.* 68, 547–556. doi: 10.1016/j.ecss.2006.03.005
- Serôdio, J., Ezequiel, J., Barnett, A., Mouget, J.-L., Méléder, V., Laviale, M., et al. (2012). Efficiency of photoprotection in microphytobenthos: role of vertical migration and the xanthophyll cycle against photoinhibition. *Aquat. Microb. Ecol.* 67, 161–175. doi: 10.3354/ame01591
- Serôdio, J., Vieira, S., Cruz, S., and Barroso, F. (2005). Short-term variability in the photosynthetic activity of microphytobenthos as detected by measuring rapid light curves using variable fluorescence. *Mar. Biol.* 146, 903–914. doi: 10.1007/s00227-004-1504-6
- Skogen Chauton, M., Winge, P., Brembu, T., Vadstein, O., and Bones, A. M. (2013). Gene regulation of carbon fixation, storage, and utilization in the diatom *Phaeodactylum tricornerutum* acclimated to light/dark cycles. *Plant Physiol.* 161, 1034–1048. doi: 10.1104/pp.112.206177
- Spilmont, N., Migné, A., Seuront, L., and Davoult, D. (2007). Short-term variability of intertidal benthic community production during emersion and the implication in annual budget calculation. *Mar. Ecol. Progr. Ser.* 333, 95–101. doi: 10.3354/meps333095
- Stock, W., Blommaert, L., Daveloose, I., Vyverman, W., and Sabbe, K. (2019). Assessing the suitability of Imaging-PAM fluorometry for monitoring growth of benthic diatoms. *J. Exp. Mar. Biol. Ecol.* 513, 35–41. doi: 10.1016/j.jembe.2019.02.003
- Underwood, G. J. C., and Kromkamp, J. C. (1999). “Primary production by phytoplankton and microphytobenthos in estuaries,” in *Advance Ecology Research*, eds D. B. Nedwell and D. G. Raffaelli (Cambridge, MA: Academic Press), 93–153. doi: 10.1016/s0065-2504(08)60192-0
- Underwood, G. J. C., Perkins, R. G., Consalvey, M. C., Hanlon, A. R. M., Oxborough, K., Baker, N. R., et al. (2005). Patterns in microphytobenthic primary productivity: species-specific variation in migratory rhythms and photosynthetic efficiency in mixed-species biofilms. *Limnol. Oceanogr.* 50, 755–767. doi: 10.4319/lo.2005.50.3.0755
- Vieira, S., Ribeiro, L., Jesus, B., Cartaxana, P., and Da Silva, J. M. (2013). Photosynthesis assessment in microphytobenthos using conventional and imaging pulse amplitude modulation fluorometry. *Photochem. Photobiol.* 89, 97–102. doi: 10.1111/j.1751-1097.2012.01224.x
- Wenderoth, K., Marquardt, J., and Rhiel, E. (2004). The big trail: many migrate at the expense of few. *Diatom Res.* 19, 115–122. doi: 10.1080/0269249x.2004.9705611
- Wenderoth, K., and Rhiel, E. (2004). Influence of light quality and gassing on the vertical migration of diatoms inhabiting the Wadden Sea. *Helgol. Mar. Res.* 58, 211–215. doi: 10.1007/s10152-004-0187-1
- Witkowski, A. (2000). Diatom flora of marine coasts I. *Iconogr. Diatomol.* 7, 1–925.

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Barnett, Méléder, Dupuy and Lavaud. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.